

DNA barcoding and phylogenetic analysis of deep-sea caridean shrimps (Decapoda: Caridea) from the southern coast of India

Rekha Devi Chakraborty*, G. Kuberan, P. Purushothaman, G. Maheswarudu, A. P. Gayathri, S. Sreelakshmy and Aghana Muralidharan
ICAR-Central Marine Fisheries Research Institute, Ernakulam North P.O., Kochi - 682 018, Kerala, India

Abstract



Knowledge on the molecular and morphological traits of deep-sea caridean shrimps from Indian waters is still sparse. This study clarifies their phylogenetic relationships and builds a DNA barcode reference using two mitochondrial markers: COI and 16S rRNA. Specimens collected during 2014-2019 across major landing centres in Kerala and Tamil Nadu yielded 15 species spanning the genera *Plesionika*, *Heterocarpus*, *Acanthephyra*, *Oplophorus*, *Parapontocaris*, *Pontocaris*, *Pasiphaea*, and *Glypocrangon*. The dataset includes 121 sequences (57 COI, 64 16S), all validated through NCBI BLAST searches. This forms the first comprehensive barcode library for deep-sea caridean shrimps from the region, supporting accurate species identification and future work on their evolution and biogeography and key for both ecological understanding and resource management.

Introduction

Caridean shrimps form one of the most diverse decapod groups, with nearly 4,000 species across 36 families occupying habitats from shallow waters to abyssal plains (De Grave and Fransen 2011; Sun *et al.*, 2024). Deep-sea carideans play key ecological roles in predation, scavenging, nutrient flux, and carbon cycling (Frolova *et al.*, 2022). Several families, particularly Pandalidae, also support commercial fisheries, although deep-sea stocks are considered vulnerable owing to slow growth rates and low reproductive output (CMFRI, 2017; FAO, 2020). Traditional taxonomy based on rostral dentition and gill traits is often confounded by convergence and subtle variation, leading to misidentifications (Gan *et al.*, 2024). Molecular markers, especially mitochondrial cytochrome oxidase subunit 1 (COI) and 16S ribosomal RNA (16S rRNA), have clarified species boundaries, revealed cryptic diversity, and reshaped caridean phylogeny (Hebert *et al.*, 2004; Li *et al.*, 2011; Liao *et al.*, 2019), showing that many families are not monophyletic (Gan *et al.*, 2024; Sun *et al.*, 2024).

In India, most deep-sea shrimp studies remain morphology-based largely, with limited molecular evidence, despite 156 species being recorded and no national DNA barcode reference library available, which remains a critical gap (George and Rao, 1966; Mohamed and Suseelan, 1973; Radhika, 2004; Karuppasamy *et al.*, 2006; Chakraborty *et al.*, 2015, 2023; Kuberan *et al.*, 2018). This study addresses this gap by integrating morphology with molecular analyses to generate deep-sea carideans' barcode dataset from the southern coast of India.

Materials and methods

Study site

Deep-sea caridean shrimps were collected between 2014 and 2019 from major commercial harbours along the southern coast of India: Kalamukku (9°59'02.91"N; 76°14'33.14"E) and Sakthikulangara (8°56'60.78"N; 76°32'34.27"E) in Kerala, and Tuticorin (8°47'40"N; 78°09'37"E) in Tamil Nadu (Fig. 1). Samples were collected

*Correspondence e-mail:
rekhadevi7674@gmail.com

Keywords:

COI gene, Deep-sea shrimps, Mitochondrial markers, Molecular identification, 16S rRNA

Received : 12.12.2024

Accepted : 12.11.2025

from deep-sea bottom trawlers operating at 200–800 m depth with a 20–26 mm cod-end mesh, trawled for 30–60 min at 3–4 knots. Specimens were immediately preserved in insulated ice boxes and transported to the laboratory for analyses.

Specimens were sorted and photo-documented. Morphological identification followed standard taxonomic references (Alcock 1901; Chace, 1985; Suseelan, 1985; Chan, 1996), ensuring accurate linkage between morphology and DNA data. Approximately 100 mg of pleopod tissue was excised from each species, preserved in 95% ethanol, and stored in pre-labelled Eppendorf tubes for molecular analysis. The integrated approach supports reliable species verification and downstream genetic studies.

Mitochondrial DNA analysis

Genomic DNA extraction, PCR amplification and sequencing

Total genomic DNA was extracted from pleopod tissue using the DNeasy® Blood and Tissue Kit (Qiagen) following the manufacturer's protocol. COI and 16S rRNA genes were amplified using universal primer sets: COI (Palumbi and Benzie, 1991): 5'-CGCCTGTTTATCAAAACAT-3'(F), 5'-CCGGTCTGAACATCACGT-3'(R); and 16S (Folmer *et al.*, 1994): 5'-CGTCAACAAATCATAAAGATATTGG-3'(F), 5'-TAAACTTCAGGGTGACCAAAAAATCA-3'(R).

PCR amplification was carried out in 25 μ l reactions containing 50 ng μ l⁻¹ genomic DNA, 0.05 U μ l⁻¹ Taq polymerase, 3 mM MgCl₂, 10 pmol of each primer, 200 μ M dNTPs and 1X PCR buffer. The cycling profile included: 94°C for 5 min; 35 cycles of 94°C for 1 min, 52°C for 1 min, and 72°C for 1.5 min; followed by a final extension at 72°C for 5 min. Amplicons were checked on 1.8% TBE agarose gels, stained with ethidium bromide and visualised

under UV. PCR products were purified with the XcelGen Gel/PCR Purification Kit and sequenced (BigDye Terminator v3.1) on an ABI Prism 3770 platform (Scigenome, India).

COI and 16S sequences were verified using ABISeq Editor v1.0 and edited in BioEdit v7.0.5.2 (Hall, 1999). Edited sequences were aligned with GenBank references using ClustalW. All sequence gaps were treated as missing data and the final sequences were submitted to GenBank. Phylogenetic analyses were performed in MEGA7 using Maximum Parsimony approach and phylogenetic trees were reconstructed applying the Subtree-Pruning-Rerooting (SPR) algorithm (Nei and Kumar, 2000), with 1,000 bootstrap replicates. Pairwise genetic distances were calculated in MEGA7. Maximum likelihood analyses were also performed for each gene, with the best-fit models selected by MEGA7: GTR+G+I for COI and TrN+G+I for 16S. ML trees were generated with 1,000 bootstrap replicates (Tamura *et al.*, 2013).

Results

This study analysed 121 mitochondrial sequences [57 COI (665 bp); 64 16S (540 bp)] from 15 deep-sea caridean shrimp species representing 8 genera and 5 families. All sequences were validated through BLAST searches and deposited in GenBank (Accession numbers given in Table 1). The species studied include *Plesionika quasigrandis*, *P. narval*, *P. semilaevis*, *P. alcocki*, *P. reflexa*, *Heterocarpus chani*, *H. woodmasoni*, *Acanthephyra fimbriata*, *A. sanguinea*, *Oplophorus gracilirostris*, *Pasiphæa alcocki*, *Parapontocaris bengalensis*, *Pontocaris affinis*, *P. propensalata*, and *Glypocrangon investigatoris* (Fig. 2).

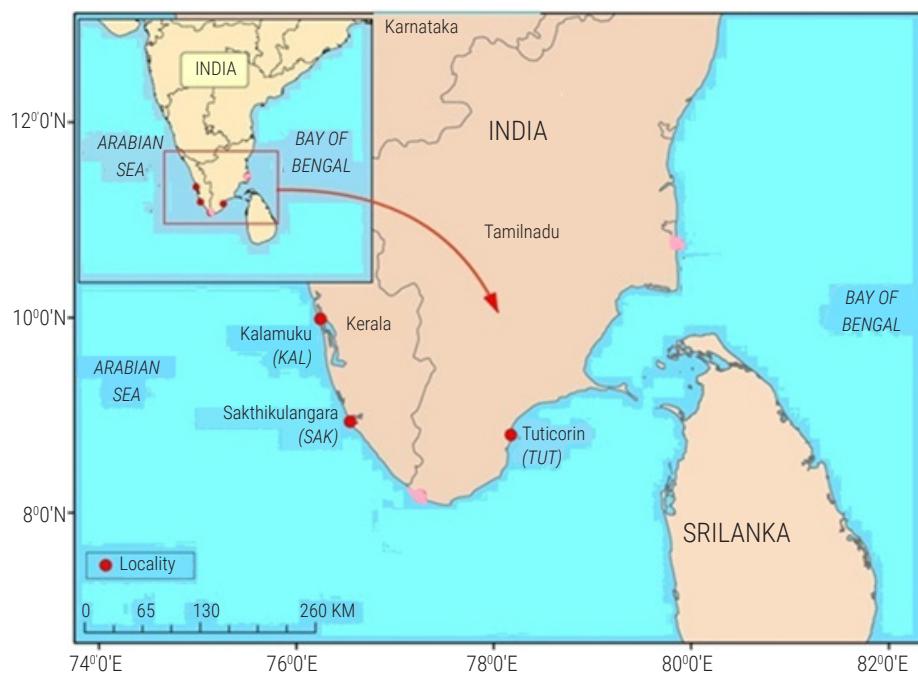


Fig. 1. Location of sampling areas along the southern coast of India

Table 1. Species, sampling locations, and GenBank accession numbers of deep-water caridean shrimps from the southern coast of India

Sl No.	Species	Sampling location	COI	16S rRNA
1	<i>Plesionika quasigrandis</i>	Sakthikulangara	KM096444	KM057395
		Sakthikulangara	KM096445	KM057396
		Sakthikulangara	KM096446	KM057397
		Sakthikulangara	KM096447	KM057398
		Sakthikulangara	KM096448	KM057399
		Sakthikulangara	KM096449	KM057400
		Sakthikulangara	KM096450	KM057401
		Sakthikulangara	KM096451	KM057402
		Sakthikulangara	KM096452	KM057403
		Sakthikulangara	KM096453	KM057404
		Sakthikulangara	KM096454	KM057405
		Sakthikulangara	KM096455	KM057406
		Sakthikulangara	KM096456	KM057407
		Sakthikulangara	KM096457	KM057408
		Sakthikulangara	KM096458	KM057409
		Sakthikulangara	KM096459	KM057410
		Kalamuku	KM096460	KM057411
		Kalamuku	KM096461	KM057412
		Kalamuku	KM096462	KM057413
		Kalamuku	KM096463	KM057414
		Kalamuku	KJ401314	KJ363166
		Kalamuku	KF938650	KJ380892
		Tuticorin	KX838917	KX838920
		Tuticorin	KX838918	KX838921
2	<i>Plesionika narval</i>	Kalamuku	KP398864	KM057378
		Kalamuku	KP398863	KP398866
		Kalamuku		KM047390
		Kalamuku		KM047389
3	<i>Plesionika semilaevis</i>	Kalamuku	KX364192	KX364190
		Kalamuku	KX364193	KX364191
4	<i>Plesionika alcocki</i>	Tuticorin	KX530799	KX364188
		Tuticorin	KX530800	
5	<i>Plesionika reflexa</i>	Sakthikulangara	MG958591	MG958587
		Sakthikulangara	MG958592	MG958588
		Sakthikulangara		MG958589
		Sakthikulangara		MG958590
6	<i>Heterocarpus chani</i>	Sakthikulangara	KX364187	KX364166
		Sakthikulangara	KX364180	KX364167
		Sakthikulangara	KX364181	KX364168
		Kalamuku	KX364182	KX364169
		Kalamuku	KX364185	KX364172
		Kalamuku	KX364186	KX364173
		Tuticorin	KX364183	KX364170
		Tuticorin	KX364184	KX364171
7	<i>Heterocarpus woodmasoni</i>	Sakthikulangara	KX364160	KX364164
		Sakthikulangara	KX364161	KX364165
		Sakthikulangara	MG879531	
		Sakthikulangara	MG958585	
		Sakthikulangara	MG958586	
		Sakthikulangara	MG879532	
		Kalamuku	MG879529	
		Kalamuku	MG879530	
		Tuticorin	KX838919	KX838922

Conntd.....

Sl no	Species	Sampling location	COI	16S rRNA
8	<i>Acanthephyra fimbriata</i>	Tuticorin	KX364162	KX364174
		Tuticorin	KX364163	KX364175
9	<i>Acanthephyra sanguinea</i>	Sakthikulangara	KU055638	KU055642
		Kalamuku	KU055639	KU055643
		Sakthikulangara	KU055644	MF627729
10	<i>Oplophorus gracilirostris</i>	Sakthikulangara	KJ472213	KJ819551
		Sakthikulangara	KJ472214	
		Sakthikulangara		
11	<i>Pasiphaea alcocki</i>	Sakthikulangara	MG748565	MG748566
12	<i>Parapontocaris bengalensis</i>	Sakthikulangara	MH045675	MH045676
13	<i>Pontacaris affinis affinis</i>	Sakthikulangara	MF996922	MF996919
		Sakthikulangara	MF996923	MF996920
		Sakthikulangara		MF996921
14	<i>Pontacaris propensalata</i>	Sakthikulangara		MF996924
		Sakthikulangara		MF996925
15	<i>Glypocrangon investigatoris</i>	Sakthikulangara	MH923239	MH923238



Fig. 2. Deep-sea shrimps from the southern coast of India (a) *Plesionika quasigrandis* (b) *Plesionika narval* (c) *Plesionika semilaevis* (d) *Plesionika alcocki* (e) *Plesionika reflexa* (f) *Heterocarpus chani* (g) *Heterocarpus woodmasoni* (h) *Acanthephyra fimbriata* (i) *Acanthephyra sanguinea* (j) *Oplophorus gracilirostris* (k) *Pasiphaea alcocki* (l) *Parapontocaris bengalensis* (m) *Pontacaris affinis affinis* (n) *Pontacaris propensalata* (o) *Glypocrangon investigatoris*

Systematics

Family Pandalidae Haworth, 1825
Genus *Plesionika* Spence Bate, 1888

Plesionika quasigrandis Chace, 1985

Material examined: 7 ovigerous females (CL 19–24 mm), 3 males (CL 18–20 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 250–300 m depth, 2 February 2014; 1 ovigerous female (CL 20 mm), Kalamukku Fishing Harbour, Kochi (9°59'02.91" N; 76°14'33.14" E), 200–300 m depth, 12 February 2014.

Distribution: The species is widely distributed across the Indo-West Pacific, from the Gulf of Aden to Japan, Southeast Asia, Australia, and India. In India it occurs along the southwest coast and the Gulf of Mannar. It inhabits the upper continental slope, typically at 200–500 m, with records ranging from 160 to 800 m depth.

Plesionika narval (Fabricius, 1787)

Material examined: 7 ovigerous females (CL 19–24 mm), 5 non-ovigerous females (CL 13–15 mm), 14 males (CL 10–15 mm), Kalamukku Fish Harbour, Kochi (9°59'02.91" N; 76°14'33.14" E), 200–300 m depth, 4 April 2014.

Distribution: The species is broadly distributed from the eastern Atlantic to the Mediterranean and occurs widely in the Indo-West Pacific, including the Red Sea, Arabian Sea, and Indian coast. It inhabits 35–400 m globally, with Indian records at 200–300 m on the southwest coast.

Plesionika semilaevis Spence Bate, 1888

Material examined: 2 ovigerous females (CL 20 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 8 February 2015; 2 males (CL 16–20 mm), Kalamukku Fishing Harbour, Kochi (9°59'02.91" N; 76°14'33.14" E), 200–300 m depth, 10 November 2014; 1 ovigerous female (CL 20 mm), 1 non-ovigerous female (CL 18 mm), 1 male (CL 17 mm), Tuticorin Fishing Harbour, Tamil Nadu (8°47'40" N; 78°09'37" E), 200–350 m depth, 20 January 2017.

Distribution: Philippines, Indonesia, South and East China Seas, Japan, and Australia. Also distributed in the southwest and southeast coast of the Indian EEZ. Depth range: 200–350 m.

Plesionika alcocki (Anderson, 1896)

Material examined: 3 males (CL 22–25 mm), Tuticorin Fishing Harbour, Tamil Nadu (8°47'40" N; 78°09'37" E), 200–300 m depth, 8 January 2015.

Distribution: Indo-West Pacific from East Africa to Japan. Found at 287–1170 m globally; in India, recorded only from the southeast coast (Bay of Bengal) at 200–300 m.

Plesionika reflexa Chace, 1985

Material examined: 1 ovigerous female (CL 15 mm) on 23 November 2015 and 1 male (CL 14.3 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), depth range 200–300 m.

Distribution: Indo-West Pacific, from the Red Sea and western Indian Ocean to the Philippines and Japan; in India, recorded from

the southwest coast (Arabian Sea). Occurs globally at 191–910 m, with Indian records at 200–300 m off Kollam.

Genus *Heterocarpus* A. Milne-Edwards, 1881

Heterocarpus chani Li, 2006

Material examined: Specimens were collected from 200–350 m depth at four sites: Sakthikulangara Fishing Harbour, Kollam (4 ovigerous females, CL 25–30 mm; 2 February 2014); Kalamukku, Kochi (3 males, CL 18–25 mm; 12 February 2014); Tuticorin Fishing Harbour (3 ovigerous females, 1 non-ovigerous female, 1 male; CL 17–25 mm; 8 January 2015); and Nagapattinam (1 ovigerous female, 1 non-ovigerous female, 1 male; CL 17–28 mm; 24 March 2017). One specimen was deposited as a voucher in the Marine Biodiversity Museum of ICAR-CMFRI, Kochi (E.D. 2.4.1.4), with the remainder retained in the working collection.

Distribution: Southern South China Sea, Bohol Sea and Sulu Sea (Philippines), at depths of 382–888 m; Bay of Bengal and Arabian Sea (India), at depths of 200–350 m.

Heterocarpus woodmasoni Alcock, 1901

Material examined: 3 ovigerous females (CL 22–28 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 18 March 2014; 3 Male (CL 20–25 mm), Kalamukku Fishing Harbour, Kochi (9°59'02.91" N; 76°14'33.14" E), 200–300 m depth, 28 March 2014; 2 ovigerous females (CL 20–25 mm), 1 non-ovigerous female (CL 18 mm), 3 males (CL 21–24 mm), Tuticorin Fishing Harbour (8°47'40" N; 78°09'37" E), 200–350 m depth, 8 January 2016.

Distribution: Northern Indian Ocean, India, and the Andaman Sea (318–485 m depth). Now from the southern coast of India (both east and west coasts) at a depth of 200–350 m.

Family Acanthephyridae Spence Bate, 1888

Genus *Acanthephyra* A. Milne-Edwards, 1881

Acanthephyra fimbriata Alcock and Anderson, 1894

Material examined: 3 males (CL 26–28 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 3 February 2015; 1 male (CL 30 mm), Kalamukku Fishing Harbour, Kochi (9°59'02.91" N; 76°14'33.14" E), 200–300 m depth, 18 November 2015.

Distribution: Gulf of Aden, Andaman Sea, Bay of Bengal, Laccadive Sea, and Arabian Sea: off Goa and the Philippines occurring between 412–1785 m. Now obtained off the Kerala coast between 200 and 350 m depths.

Acanthephyra sanguinea Wood-Mason and Alcock, 1892

Material examined: 2 ovigerous females (CL 23–25 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 18 March 2014; 3 males (CL 18–21 mm), Kalamukku Fishing Harbour, Kochi (9°59'02.91" N; 76°14'33.14" E), 200–300 m depth, 28 March 2014; 1 ovigerous female (CL 25 mm), 1 non-ovigerous female (CL 18 mm), 1 male (CL 17 mm), Tuticorin Fishing Harbour (8°47'40" N; 78°09'37" E), 200–350 m depth, 29 January 2017.

Distribution: Indo-West Pacific; Gulf of Aden and Eastern African coast, depth 3200 m. Now southern coast of India is at a depth of 200–350 m.

Family Oplophoridae Dana, 1852

Genus *Oplophorus* H. Milne Edwards, 1837

Oplophorus gracilirostris A. Milne Edwards, 1881

Material examined: 6 non-ovigerous females (CL 16–18 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 6 January 2014; 3 males (CL 20–25 mm); 13 non-ovigerous females (CL 17–20 mm), Tuticorin Fishing Harbour (8°47'40" N; 78°09'37" E), 400–800 m depth, 24 January 2014.

Distribution: Off south-eastern Africa, the Indian Ocean, Indonesia, the Philippines, southern Japan, Fiji Islands, Hawaii, the Gulf of Mexico, the Bahamas and Caribbean Sea. The specimen in the present study was from a depth of 400–800 m along the southern coast of India.

Family Pasiphaeidae Dana, 1852

Genus *Pasiphaea* Savigny, 1816

Pasiphaea alcocki Wood-Mason and Alcock, 1891

Material examined: 3 males (CL 20–23 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 16 November 2013.

Distribution: The species is widespread in the Indo-West Pacific and southwestern Atlantic; in India it occurs in the Arabian Sea and Bay of Bengal. Previously recorded from 335–1732 m, this study reports *P. alcocki* off Sakthikulangara at a shallower depth of 200–300 m.

Family Crangonidae Haworth, 1825

Genus *Parapontocaris* Alcocki, 1901

Parapontocaris bengalensis (Wood-Mason and Alcock 1891)

Material examined: 3 ovigerous females (CL 9–10 mm), 1 male (CL 9 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 12 October 2013.

Genus *Pontocaris* Spence Bate, 1888

Pontocaris affinis affinis (Alcock, 1901)

Material examined: 20 non-ovigerous females (CL 9–11 mm), 18 males (CL 10–11 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 16 September 2014.

Distribution: Along the Indian coast, *P. affinis affinis* was previously reported from Bombay, the Bay of Bengal, and Madagascar at 33–175 m depth. This first record from the Quilon Bank at 250–300 m indicates a much wider depth range in the Indian Ocean.

Pontocaris propensalata Spence Bate, 1888

Material examined: 2 non-ovigerous females (CL 10–11 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 10 December 2015.

Distribution: Distributed across the Andaman Sea, Philippines, Indonesia, and the south-west Pacific at depths of 100–525 m. The present specimens were collected at 250–300 m on the continental slope of the south-eastern Arabian Sea.

Family Glyphocrangonidae Smith, 1884

Genus *Glypocrangon* A. Milne-Edwards, 1881

Glypocrangon investigatoris Wood-Mason and Alcock, 1891

Material examined: 3 ovigerous females (CL 26–28 mm), Sakthikulangara Fishing Harbour, Kollam (8°56'60.78" N; 76°32'34.27" E), 200–300 m depth, 18 December 2017.

Distribution: Bay of Bengal, Arabian Sea, Myanmar, Sri Lanka; Depth 145–803 m.

A total of 57 COI sequences representing 14 deep-sea caridean shrimp species collected from 200–800 m depth along the southern coast of India were generated and deposited in GenBank. Sequence lengths ranged from 451–684 bp, yielding 17 haplotypes with high haplotype diversity (Hd = 0.82). The dataset showed an overall A+T bias (57.3%), highest at the third codon position (62.9%). Mean interspecific genetic distance across taxa was 26%, with maximum divergence (43%) between *P. quasigrandis* and *O. gracilirostris*, and minimum divergence (21%) between *Plesionika reflexa* and *H. chani*. Phylogenetic relationships were inferred using Maximum Likelihood analysis (GTR+G+I, 1000 bootstraps) (Fig. 3).

Within *Plesionika*, five species were identified from 29 sequences, with a mean interspecific divergence of 13% and no intraspecific variation; notably, *P. quasigrandis* showed complete genetic homogeneity across the Arabian Sea and Bay of Bengal, confirming earlier misidentifications. Two *Heterocarpus* species exhibited low intrageneric divergence (1%) and moderate interspecific divergence (24%), while two *Acanthephyra* species showed 20% divergence. Several rare or data-deficient taxa (*O. gracilirostris*, *Pontocaris affinis affinis*, *Parapontocaris bengalensis*, *Pasiphaea alcocki*, and *Glypocrangon investigatoris*) were also barcoded, all lacking intraspecific divergence, though high interspecific divergence (31%) was observed between *P. affinis affinis* and *P. bengalensis*.

In 16S rRNA analyses, sequence lengths varied from 314 bp in *Oplophorus gracilirostris* to 671 bp in *G. investigatoris*, followed by *P. reflexa* (650 bp) and *Acanthephyra sanguinea* (628 bp). Multiple sequence alignment (BioEdit) produced a consensus length of 655 sites, including 61 variable sites (54 parsimony-informative, 7 singletons) and 34 invariable sites, with 465 gaps or missing data. Twenty haplotypes were identified. Mean nucleotide composition of the 16S rRNA dataset (n = 64) was A 32.2%, G 22.3%, C 11.0%, and T 34.5%. Mean pairwise K2P genetic distance was 31%, with maximum divergence between *P. narval* and *P. propensalata* (59%) and minimum between *P. propensalata* and *Pontocaris affinis affinis* (9%). Phylogenetic relationships inferred using Maximum Likelihood (GTR+G+I, 1000 bootstraps) are shown in Fig. 4.

Pandalidae: Five *Plesionika* species (*P. quasigrandis*, *P. narval*, *P. alcocki*, *P. reflexa*, and *P. semilaevis*; n = 33) were analysed, with sequence lengths ranging from 484 to 644 bp. Alignments yielded 525 conserved and 53 variable sites (48 parsimony-informative). Mean nucleotide composition was A 32.7%, G 23.3%, C 11.2%, and T 32.8% (A+T = 65.5%). Mean genetic distance within *Plesionika* was 11%, with maximum divergence between *P. narval* and *P. semilaevis* (32%) and minimum between *P. reflexa* and *P. semilaevis* (15%). Intraspecific divergence was low (0–1%).

Two *Heterocarpus* species (*H. chani* n=8; *H. woodmasoni* n=5) were analysed using 16S rRNA (376 bp). Of 367 sites, 35 were parsimony-informative. Mean nucleotide composition showed

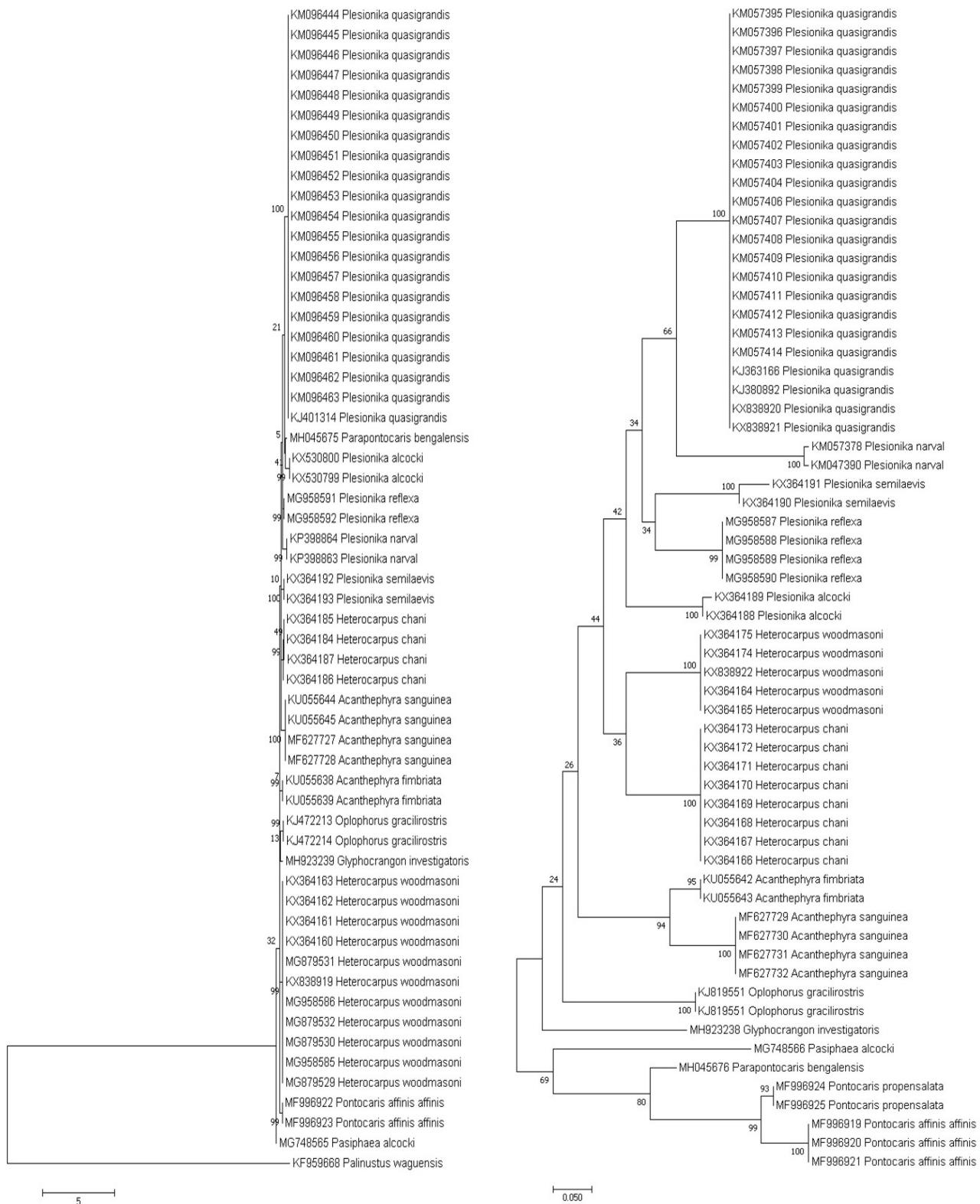


Fig. 3. Phylogenetic tree of the identified caridean shrimps using Maximum-Likelihood with 1000 bootstraps under the best fitting model GTR+G+I inferred from DNA Sequences of mitochondrial gene COI; Samples collected from the southern coast of India

Fig. 4. Phylogenetic tree of the caridean shrimps from the southern coast of India using Maximum-Likelihood with 1000 bootstraps under the best fitting model GTR+G+I inferred from DNA sequences of mitochondrial 16S rRNA gene

strong A+T bias (68.7%). Mean genetic distance was 5%, with 10% interspecific divergence and no intraspecific variation across regions.

Acanthephyridae: The family Acanthephyridae was represented by two *Acanthephyra* species, *A. fimbriata* (n=2) and *A. sanguinea* (n=4), analysed using 16S rRNA sequences (485 bp). Of the 344 aligned sites, 315 were invariable and 29 were parsimony-informative, with no singleton sites. Mean nucleotide composition was A 29.6%, G 22.5%, C 10.9%, and T 37.4% (A+T = 67%), increasing to 72.6% at the third codon position. The mean genetic distance within *Acanthephyra* was 5%, with an interspecific K2P distance of 9% and no intraspecific divergence detected in either species.

Crangonidae: Within Crangonidae, two genera were analysed: *Parapontocaris* (*P. bengalensis*, n=1) and *Pontocaris* (*P. affinis affinis* and *P. propensalata*). The 16S sequences ranged from 481 to 493 bp. Alignment yielded 469 sites, including 61 variable and 17 parsimony-informative sites. Mean nucleotide composition was A 30.5%, G 23.9%, C 11.8%, and T 33.8% (A+T = 64.3%), increasing to 66.6% at the third codon position. The mean genetic distance among species was 6%, with interspecific K2P distances ranging from 4% to 13%. The lowest divergence (4%) occurred between *P. propensalata* and *P. affinis affinis*, while no intraspecific divergence was detected in either species.

Other carideans: We also recorded three other caridean species, *O. gracilirostris* (n=2) from the family Oplophoridae, *G. investigatoris* (n=1) from the family Glyphocrangidae, and *P. alcocki* (n=1) from the family Pasiphaeidae. All of these species were collected from the south-west coast of India. For the COI gene, we observed amplified sequence lengths of 284 bp for *O. gracilirostris*, 540 bp for *G. investigatoris*, and 510 bp for *P. alcocki*. Based on the results of genetic analysis, we found that the intraspecific genetic distance for the specimens of *O. gracilirostris* was 0%, indicating a lack of genetic variation among these individuals.

Discussion

Taxonomic resolution of deep-sea shrimps is frequently constrained by morphological complexity and convergence, necessitating the use of molecular tools for accurate species identification (Gomon et al., 2014). DNA barcoding, which relies on the principle that interspecific genetic divergence exceeds intraspecific variation, has proven to be a rapid and reliable approach for species delimitation across animal taxa, including crustaceans (Hebert et al., 2004; Goldstein and DeSalle, 2011). Deep-sea caridean shrimps remain among the least explored crustacean groups, with persistent taxonomic uncertainties across families, although previous studies employing mitochondrial, nuclear ribosomal, and protein-coding genes have improved understanding of higher-level relationships and species complexes within Carideans (Li et al., 2011). In this context, the present study provides a substantial contribution to deep-sea caridean shrimp diversity along the southern coast of India by integrating COI and 16S rRNA markers, generating COI barcodes for 14 species and 16S sequences for 15 species, and confirming 11 new regional records through a combination of morphological and molecular evidences.

This study provides robust molecular evidence for accurate species delimitation within the genus *Plesionika* from Indian waters. Phylogenetic analyses based on COI and 16S rRNA genes clearly resolved five well-supported clades in this genus: *P. quasigrandis*, *P. narval*, *P. semilaevis*, *P. alcocki*, and *P. reflexa*. Molecular comparison of Indian *P. quasigrandis* with the Philippine type material showed COI divergence of 5.8–8.4%, confirming its identity and correcting earlier misidentification as *P. spinipes* in Indian waters (Chakraborty et al., 2015). Specimens previously identified as *P. martia* from multiple deep-sea landing centres along the southern Indian coast were genetically and morphologically confirmed as *P. semilaevis*. Comparative analyses with GenBank sequences showed substantial divergence between *P. semilaevis* and *P. martia*, particularly in COI (up to 26.1%), exceeding the commonly accepted crustacean species threshold (>3%), conclusively establishing *P. semilaevis* as the species occurring in Indian waters.

Genetic comparisons of Indian *P. reflexa* with topotypic material of *P. ensis* and *P. reflexa* revealed high inter-regional COI divergence (9.3–14.5%), while divergence within Indian material remained low (0–0.3%). Despite limited genetic differentiation, consistent morphological differences, especially reduced or absent epipods on pereiopods III and IV, suggest that the Indian form may represent a distinct taxon (Chan et al., 2018). *Plesionika narval* from the south-west coast of India clustered distinctly in ML analyses and showed moderate genetic divergence from closely related species (*P. grandis*, *P. ensis*, *P. spinipes*, and *P. williamsi*). For *P. alcocki*, COI and 16S sequences showed no intraspecific divergence, and strong bootstrap support confirmed its stable presence along the Indian coast.

The genus *Heterocarpus* comprises commercially important deep-sea shrimps with a global diversity of about 30 species (Holthuis, 1980). This study reports the first COI and 16S rRNA sequences of *H. chani* and *H. woodmasoni* from Indian waters. *Heterocarpus chani*, previously misidentified as *H. gibbosus* in India, was earlier confirmed morphologically, and its COI and 16S sequences generated here were deposited in GenBank. Comparative analysis showed low intraspecific divergence in *H. chani* (0–2% for COI; 0% for 16S), while interspecific divergence from the closely related *H. gibbosus* was higher (12–13% for COI; 0–1% for 16S), providing molecular support for its occurrence in Indian waters. Similarly, *H. woodmasoni*, another commercially important species, was sequenced for COI and 16S and compared with available GenBank data. Low to moderate intraspecific divergence was observed (0–9% for COI; 0–4% for 16S), further validating its taxonomic identity in Indian waters.

This study analysed COI and 16S rRNA sequences of two *Acanthephyra* species (*A. fimbriata* and *A. sanguinea*) from Indian waters. *Acanthephyra fimbriata*, which shows morphological resemblance to *A. armata*, was compared with GenBank sequences. Intraspecific divergence between Indian and Philippine material (COI: KP076185; 16S: KP075895) was low (3% for COI; 0.3% for 16S), while Indian specimens showed complete genetic homogeneity (COI: KT222917, KT222918, MG029405; 16S: KP372713). Interspecific divergence from other *Acanthephyra* species ranged from 17.5–20.9% (COI) and 5.2–9.5% (16S), consistent with earlier findings.

A. sanguinea, commonly occurring along the southern coast of India, is reported here for the first time using molecular data, with sequences deposited in GenBank. Phylogenetic analyses based on COI and 16S showed no intraspecific divergence among Indian

specimens. Comparisons with congeners revealed interspecific divergence of 7.4% from *A. fimbriata* (KU055642, KU055643) using 16S (KU055642, KU055643) and 22.9% using COI (KU055644, MF627727), supporting clear species-level separation.

In this study, we present the first report of the species *O. gracilirostris* from India, along with its molecular data (COI and 16S). To determine the genetic relationships and phylogeny, pairwise genetic distance and sequence identity analysis of COI and 16S genes were conducted, focusing on closely related species, namely *O. typus* and *O. spinosus*. Comparing COI sequences with *O. typus* and *O. spinosus* showed interspecies genetic distances of 0.020 (2%) and 0.131 (13%), respectively. Similarly, comparing 16S DNA sequences of *O. gracilirostris* available in NCBI GenBank with the present sequence of *O. gracilirostris* showed an intraspecies genetic distance of 0.03 (3%) and interspecies distance of 0.05 (5%) with *O. typus* (Chakraborty *et al.*, 2014).

The genus *Parapontocaris* comprises six species with a global distribution. In this study, we focused on *P. bengalensis* for which mitochondrial gene sequencing of COI and 16S rDNA was performed and these sequences were subsequently deposited in GenBank. The sequence lengths for COI and 16S genes were determined to be 634 and 481 bp respectively. To assess the genetic relationships, we conducted interspecies divergence analysis, comparing the COI and 16S sequences from the present *P. bengalensis* specimen with other sequences retrieved from the NCBI database. The results revealed substantial genetic differences between *P. bengalensis* and *P. levigata* (KP759476, 9.1%) as well as *Parapontocaris* sp. (Accession No. KP759477, 8.8%) when analysing COI sequences. On the other hand, for 16S sequences, the genetic differences were observed to be in the range of 2.1–6.2% with *P. levigata*, *Parapontocaris* sp., and *P. aspera*.

Mitochondrial gene sequencing was conducted for *P. affinis affinis*, focusing on COI, with accession numbers MF996922 and MF996923, and for 16S rDNA with accession numbers MF996919, MF996920, and MF996921. Additionally, the 16S rDNA sequences of *P. propensalata* were analysed, and their accession numbers are MF996924 and MF996925. Upon comparing the 16S rDNA sequences, a genetic divergence of 4.1% was observed between *P. affinis affinis* and *P. propensalata* (Purushothaman *et al.*, 2019).

We retrieved COI and 16S sequences of twenty-two and nine species, respectively, from the NCBI database, including one intraspecies sequence of *P. alcocki*. For the COI gene, the following species were analysed: *P. sivado* (KP759488 and JQ306265), *P. telacantha* (KP759492), *P. multidentata* (JQ305978, FJ581855, and KF931036), *P. planidorsalis* (KP759483), *P. sirenkoi* (KP759484), *P. tarda* (DQ882139, JQ305981, and AF125439), *P. merriami* (MF197272), *P. pacifica* (DQ882135), and *P. hoplocerca* (JQ306169). For the 16S gene, the following species were considered: *P. sivado* (MF279526 and KP725631), *P. telacantha* (KP725635), *P. multidentata* (MF279519), *P. planidorsalis* (KP725624), *P. levicarinata* (MF279517 and GQ131899), *P. japonica* (MF279516), *P. sirenkoi* (MF279525 and KP725625), *P. merriami* (MF197216 and EU868700), *P. mclaughlinae* (MF279518), *P. americana* (MF279511), *P. acutifrons* (MF279508), *P. sinensis* (MF279524), *P. diaphana* (MF279512), *P. aequus* (MF279509), *P. falx* (MF279514), *P. romenskyi* (MF279522), *P. gelasinus* (MF279515), *P. scotiae* (MF279523), *P. orientalis* (MF279520), *P. pseudacantha* (MF279521), and *P. exilimanus* (MF279513). Interspecific divergence was higher for COI (18.5–27.9%) than for 16S (7.4–26.4%). Although

genetic divergence between *P. sirenkoi* and the present *P. alcocki* isolate was relatively low (16S: 7.4%; COI: 18.5%), clear morphological differences were observed. The present specimen bears 10–11 spines on the merus of the second pereiopod, compared to a single spine in *P. sirenkoi*, supporting the substantial intraspecific divergence (21.2%) observed within *P. alcocki* (16S: MF279510). Mitochondrial COI and 16S rRNA sequences of *Glypocrangon investigatoris* (618 bp and 464 bp, respectively) were generated and deposited in GenBank. Intraspecific comparison of COI sequences with available NCBI data showed low divergence (0.6–0.7%) from *G. investigatoris* (KJ143751, KJ143752). Comparable 16S data for this species were unavailable in GenBank. Interspecific comparisons with other *Glypocrangon* species revealed higher divergence in COI (5.1–26.5%) than in 16S (1.1–8.1%). Among congeners, *G. regalis* showed the lowest divergence from the present material (COI: ~5%; 16S: 1.1%) (Kuberan *et al.*, 2019).

The present study demonstrates the pivotal role of mitochondrial DNA (mtDNA) divergence in accurately identifying commercially important deep-sea shrimp species. The results highlight the effectiveness of molecular approaches in resolving species boundaries and clarifying phylogenetic relationships. Consistently lower intraspecific than interspecific genetic distances confirm the strong diagnostic value of mtDNA markers for distinguishing closely related shrimp species.

Acknowledgements

The authors thank the Science and Engineering Research Board (SERB), Department of Science and Technology, Government of India, for financial support, and the Director, ICAR-CMFRI, Kochi for encouragement and support.

References

- Alcock, A. 1901. Catalogue of the Indian decapod crustacea in the collection of the Indian Museum, Vol. 7. Indian Museum, Kolkata, India. <https://doi.org/10.5962/bhl.title.10323>.
- Alcock, A. and Anderson, A. R. 1894. Natural history notes from H.M. Indian marine survey steamer *Investigator*. Series II, No. 17. *J. Asiat. Soc. Bengal*, 63(2): 197–209.
- Chace, F.A. Jr. 1985. The caridean shrimps (Crustacea: Decapoda) of the Albatross Philippine Expedition, 1907–1910, Part III: Families Thalassocarididae and Pandalidae. *Smithson. Contrib. Zool.*, 411: 1–143. <https://doi.org/10.5479/si.00810282.411>.
- Chakraborty, R. D., Chan, T. Y., Maheswarudu, G., Kuberan, G., Purushothaman, P., Chang, S. C. and Jomon, S. 2015. On *Plesionika quasigrandis* Chace, 1985 (Decapoda: Caridea: Pandalidae) from southwestern India. *Crustaceana*, 88(7–8): 923–930. <https://doi.org/10.1163/15685403-00003451>.
- Chakraborty, R. D., Purushothaman, P., Kuberan, G., Sebastian, J. and Maheswarudu, G. 2014. Identification of *Oplophorus gracilirostris* from the southeast and southwest coast of India using mitochondrial genetic marker. *Int. J. Sci. Res.*, 3(6): 167–169. <https://www.doi.org/10.36106/ijsr>.
- Chan, T. Y. 1996. Crustacea Decapoda Crangonidae: Revision of *Aegaeon*, *Pontocaris* and *Parapontocaris*. *Memoirs of the National Museum of Natural History*, 168: 269–336 (In French).
- Chan, T. Y., Chakraborty, R. D., Purushothaman, P., Kuberan, G. and Yang, C. H. 2018. On *Plesionika persica* and *P. reflexa* (Decapoda: Pandalidae) from India. *Zootaxa*, 4382(3): 583–591. <https://doi.org/10.11646/zootaxa.4382.3.9>.
- CMFRI 2017. CMFRI Annual report 2016–2017. ICAR-Central Marine Fisheries Research Institute, Kochi, India.

De Grave, S. and Fransen, C. H. J. M. 2011. *Carideorum catalogus*: The recent species of dendrobranchiate, stenopodidean, procarididean and caridean shrimps. Naturalis Biodiversity Center, Leiden, The Netherlands

FAO 2020. *The state of world fisheries and aquaculture 2020*. Food and Agriculture Organization of the United Nations, Rome, Italy.

Frolova, P., Horka, I. and Duris, Z. 2022. Molecular phylogeny and historical biogeography of marine palaemonid shrimps. *Sci. Rep.*, 12: 15237. <https://doi.org/10.1038/s41598-022-19460-1>.

Gan, Z., Chan, T. Y. and Chu, K. H. 2024. Integrative taxonomy reveals deep-sea caridean shrimps from the South China Sea. *Front. Mar. Sci.*, 11:1398599. <https://doi.org/10.3389/fmars.2024.1398599>.

George, M. J. and Rao, P. V. 1966. On some new records of deep-sea prawns (Decapoda: Pandalidae) from the Indian region. *Mar. Biol. Assoc. India*, 8: 142–149.

Goldstein, P. Z. and DeSalle, R. 2011. Integrating DNA barcode data and taxonomic practice. *BioEssays*, 33(2): 135–147. <https://doi.org/10.1002/bies.201000101>.

Gomon, M. F., Ward, R. D., Chapple, S. and Hale, J. M. 2014. DNA barcode evidence for species of *Chlorophthalmus*. *Mar. Freshw. Res.*, 65: 1027–1034. <https://doi.org/10.1071/MF13247>.

Hall, T. A. 1999. BioEdit: A user-friendly biological sequence alignment editor. *Nucleic Acids Symp. Ser.*, 41: 95–98.

Hebert, P. D. N., Stoeckle, M. Y., Zemlak, T. S. and Francis, C. M. 2004. Identification of birds through DNA barcodes. *PLoS Biol.*, 2(10): e312. <https://doi.org/10.1371/journal.pbio.0020312>.

Holthuis, L. B. 1980. FAO species catalogue. Vol. 1. Shrimps and prawns of the world. *FAO Fisheries Synopsis No. 125*. Food and Agriculture Organization of the United Nations, Rome, Italy.

Karuppasamy, P. K., Menon, N. G., Nair, K. K. C. and Achuthankutty, C. T. 2006. Distribution and abundance of pelagic shrimps from the deep scattering layer of the eastern Arabian Sea. *J. Shellfish Res.*, 25(3): 1013–1019.

Kuberan, G., Chakraborty, R. D., Purushothaman, P. and Maheswarudu, G. 2018. First record of *Acanthephyra fimbriata* from the southwest coast of India. *Zootaxa*, 4531(2): 288–294. <https://doi.org/10.11646/zootaxa.4531.2.7>.

Kuberan, G., Chakraborty, R. D., Purushothaman, P., Maheswarudu, G., Sreesanth, L. and Ragesh, N. 2019. New record of *Glypocrangon investigatoris* from the southeastern Arabian Sea. *Zootaxa*, 4612(4): 566–570. <https://doi.org/10.11646/zootaxa.4612.4.9>.

Li, C. P., De Grave, S., Chan, T. Y., Lei, H. C. and Chu, K. H. 2011. Molecular systematics of caridean shrimps based on five nuclear genes. *Zoologischer Anzeiger*, 250: 270–279. <https://doi.org/10.1016/j.jcz.2011.07.001> (In German).

Liao, Y., Ma, K. Y., De Grave, S., Komai, T., Chan, T. Y. and Chu, K. H. 2019. Systematic analysis of the shrimp superfamily Pandaloidea. *Mol. Phylogenet. Evol.*, 134: 200–210. <https://doi.org/10.1016/j.ympev.2019.03.018>.

Mohamed, K. H. and Suseelan, C. 1973. Deep-sea prawn resources of the southwest coast of India. *Proceedings of the Symposium on Living Resources of the Seas around India*, ICAR-Central Marine Fisheries Research Institute, pp. 614–633.

Nei, M. and Kumar, S. 2000. *Molecular evolution and phylogenetics*. Oxford University Press, New York, USA.

Palumbi, S. R. and Benzie, J. 1991. Large mitochondrial DNA differences between morphologically similar penaeid shrimp. *Mol. Mar. Biol. Biotechnol.*, 1: 27–34.

Purushothaman, P., Chakraborty, R. D., Kuberan, G. and Maheswarudu, G. 2019. Integrative taxonomy of commercially important deep-water shrimps from India. *J. Genet.*, 98: 1–13. <https://doi.org/10.1007/s12041-018-1036-7>.

Radhika, R. 2004. *Systematics, fishery and bionomics of deep-sea prawns off Kerala*. Ph. D. Thesis, Cochin University of Science and Technology, Kochi, India.

Sun, X., Chan, T. Y. and Chu, K. H. 2024. Phylogenomics and evolutionary history of caridean shrimps. *Mol. Phylogenet. Evol.*, 190: 107900. <https://doi.org/10.1016/j.ympev.2024.107900>.

Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. 2013. MEGA6: Molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.*, 30(12): 2725–2729. <https://doi.org/10.1093/molbev/mst197>.