The bloom of invasive alien comb jelly *Beroe ovata* Bruguière, 1789 in the Bay of Bengal

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The present study documented a bloom of the comb jelly *Beroe ovata* occurring off Devaneri Kuppam in Mahabalipuram, close to Chennai. The abundance of the *B. ovata* bloom was estimated as 342 ± 21 individuals per cubic metre. A negligible number of hydromedusae specimens mixed with the comb jelly were also observed. The abundance of its predatory fish, the black pomfret (*Parastromateus niger*), the hydrozoan jellyfish (*Ae-quorea pensilis*), and the scyphozoan jellyfish (*Cyanea nozakii*) would be positively correlated with the sudden disappearance of the bloom of *B. ovata* on 10 October 2024. Following the bloom of *B. ovata*, the abundance of *A. pensilis* and *C. nozakii* led to a depletion of copepods, the main predators of dinoflagellates and diatoms, favouring the bloom of *Noctiluca scintillans*.

Keywords: *Beroe ovata*, comb jelly bloom, *Cyanea nozakii*, hydromedusae, *Noctiluca scintillans*.

CTENOPHORES are gelatinous zooplankton belonging to the phylum Ctenophora found throughout the world's oceans. They are often called 'comb jellies' due to their transparent, soft, delicate and gelatinous bodies with eight comb rows composed of thousands of cilia for movement¹. To capture prey, ctenophores possess non-stinging 'colloblast' cells². The stinging cells known as 'cnidoblasts' are found in most scyphozoan jellyfish, also known as real jellyfish (Phylum Cnidaria). Ctenophores are divided into 48 genera, 33 families and 9 orders, with 185 extant species and 14 extinct species recognized worldwide. Among these, order Beroida Eschscholtz, 1825, comprises 30 species under 2 genera and 1 family³. To date, 19 ctenophores belonging to 7 families and 5 orders have been reported from India⁴. Because of distortion and shrinking, it is particularly challenging to identify preserved specimens. The majority of live specimens are harmed when they are caught in a zooplankton net. The genus Beroe has a peculiar character among other ctenophores. It lacks tentacles and feeds exclusively on other ctenophores and mesozooplankton⁵. It closes its mouth using adhesive cells (colloblasts) while moving forward or swimming⁶.

Beroe ovata Bruguière, 1789, also known as the comb jelly, is indigenous to the western Atlantic coastline, extending from Argentina to the United States. In 1997, ship ballast waters brought it to the Black Sea, from where it spread to nearby European seas^{7,8}. The occurrence of B. ovata has been documented in 16 countries worldwide to date⁹. B. ovata has an oval body that is often pink and has a broader oral end. The meridian canals with anastomoses between them are the primary identifying characteristics of B. $ovata^{10}$. Like other ctenophores, B. ovata is a hermaphrodite that can develop into larvae directly, mature quickly, have better fertility and self-fertilize. It can quick reproduce a few individuals can produce a mass population within a month under favourable conditions¹¹. Compared to the eggs of ctenophores, the bioluminescent amplitude of *B. ovata* larvae was eight times greater¹². Complete regenerative powers allow the comb jelly, Mnemiopsis leidvi, to reverse age and return to larval stag es^{13} . Nevertheless, *B. ovata* is restricted to lip development and lacks the capacity to regenerate the entire $body^{14}$.

The occurrence of ctenophore swarms or blooms in Indian seas has been reported since 1940. From September 1940 to August 1943, scientists studied plankton at Krusadai Island in the Gulf of Mannar. They found a lot of ctenophore species Pleurobrachia globosa Moser, 1903 and hydromedusae from November to February¹⁵. The bloom of P. globosa occurred on the Madaras (Chennai) coast during April-May 1962 (ref. 16). Outbreaks of ctenophore and medusae were reported at Sassoon Dock in Mumbai on 27 October 1959 and linked to a sudden rise in catches of the fish Silver pomfret, Pampus argentius¹⁷. Swarms of ctenophore P. globosa and Sagitta enflata (Chaetognatha) were reported on the Goa coast (Arabian Sea) on 15 October 1980 (ref. 18). The pH and dissolved oxygen (DO) in the ocean surrounding Port Blair in the Andaman Islands were directly correlated with the quantity of the ctenophore Pleurobrachia pileus (O. F. Müller, 1776)¹⁹. In January 2018, Purushothaman et al.²⁰ reported a swarm of P. pileus in the Sundarban coastal waters of the Bay of Bengal. In July 2019, researchers in India reported the bloom of the invasive alien ctenophore B. ovata Bruguière, 1789, in the coastal areas of Kollam, Kerala, in the Arabian Sea²¹.

The present study documented a bloom of the comb jelly B. ovata that occurred off Devaneri Kuppam in Mahabalipuram, close to Chennai. On 7 and 8 October 2024, researchers spotted a bloom in the Bay of Bengal at depths between 6 and 150 m (12°64.803'N, 80°26.352'E). Between 7 and 10 a.m., specimens were collected using a zooplankton net with a 200 µm mesh and a 60 cm diameter mouth dragged horizontally at a depth of 1 m in the sea surface area. The flow meter data facilitated the conversion of B. ovata counts into quantities per unit volume. The abundance of the B. ovata bloom was estimated as 342 ± 21 individuals per cubic metre. Among these specimens, 21% measured less than 10 mm in length, 60.5% measured between 10 and 20 mm, 11.5% measured between 20 and 40 mm, and 7% measured between 41 and 50 mm. It has been reported that B. ovata up to 8.0 mm in length were considered larva, 9-40 mm juvenile, 41-50 mm mature and greater than 50 mm adult²². As a result,

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only 7% of B. ovata were in the mature stage, ready for reproduction and 93% of specimens in the bloom were juveniles. We observed a negligible number of hydromedusae specimens mixed with the comb jellies. Many comb jellies and hydromedusae specimens suffered damage, with some losing their body parts in the zooplankton net collection. Therefore, we randomly collected live specimens using a plastic bucket. We brought approximately 46 live specimens to the laboratory and photographed them (Figure 1). We captured images of three distinct *B. ovata* sizes, shown in Figure 2 a-c. We identified the hydromedusae species as Eirene ceylonensis Browne, 1905 (Figure 2 d), Zygocanna buitendijki Stiasny, 1928 (Figure 2 e) and Aequorea pensilis (Haeckel, 1879) (Figure 2 e). Environmental parameters were also collected during the survey: sea-surface temperature (SST), 30.15°C; salinity, 33.85‰; DO, 9.34 mg/l; chlorophyll a, 0.37 mg/m³; nitrate, 0.002 mg/l; phosphate, 0.005 mg/l and silicate, 0.05 mg/l. The bloom of B. ovata completely disappeared on 10 October 2024. Instead, the

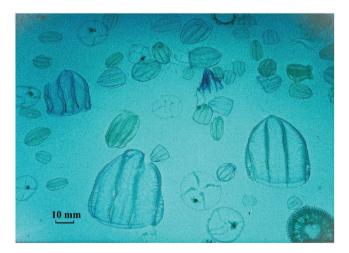


Figure 1. Comb jelly *Beroe ovata* Bruguière, 1789 collected from Bay of Bengal.

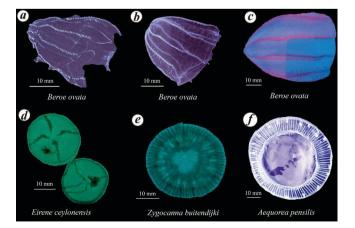


Figure 2. B. ovata comb jelly in three different sizes with morphological variation (a-c) and species of hydromedusae (d-f).

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scyphozoan jellyfish *Cyanea nozakii* Kishinouye, 1891, with a diameter ranging from 28 to 52 cm, began to appear in the sea.

The primary determinants of jellyfish and ctenophores bloom are temperature and salinity. Bloom formation is also influenced by a mix of biological and physical variables, food availability and predation pressures²³. Ctenophores digested food more quickly when the temperature and salinity rose²⁴. Salinity and temperature were recorded as 34‰ and 30°C respectively, when the bloom of P. globosa occurred in Chennai during April-May 1962 (ref. 16). Similarly, the bloom of B. ovata was caused by an unusual increase in air temperature to 33°C on 4 October, which coincided with a sudden rise in SST and salinity. The comb jelly *B. ovata* is a voracious feeder, with a daily intake of food requirements of more than 20% of its body weight²⁵. It mainly feeds on other ctenophores, pelagic fish eggs and larvae, copepods and amphipods. In turn, ctenophores are mostly consumed by fish, jellyfish, seabirds and turtles²⁶. In the present study, we hypothesized that the abrupt disappearance of the ctenophore *B. ovata* would have a positive correlation with the abundance of their predatory fish - black pomfret Parastromateus niger, hydrozoan jellyfish A. pensilis, and scyphozoan jellyfish C. nozakii. During the field study, multiple schools of black pomfrets, P. niger, were found at a depth of 4 m in the B. ovata bloom region. The feeding habits and food of pomfrets in the Bay of Bengal were thoroughly studied by researchers. Gut content analysis showed that ctenophores and medusae ranked second in the Chinese pomfret, Pampus chinensis, and third in the black pomfret, P. niger^{27,28}.

The hydrozoan jellyfish (Aequorea spp.) is known to consume a wide variety of copepods, ctenophores, siphonophores, salps, fish eggs and larvae²⁹. A high density of hydromedusae with 500 individuals/1000 m³ was recorded off Madras³⁰. Generally, Cyanea spp. consumes copepods and gelatinous zooplankton, such as ctenophores, hydromedusae and other small scyphozoan medusae. All the specimens of *B. ovata* would have been consumed by *C.* nozakii and A. pensilis. Moreover, for reproduction, ctenophores require more than 0.5 mg/m^3 of chlorophyll³¹. However, there was little chlorophyll *a* in the bloom area, which would have stopped adult B. ovata from reproducing. On 9 October 2024, we observed a large gathering of ghost jellyfish (C. nozakii) along the coast of Tamil Nadu, extending from North Chennai to Rameswaram. Numerous fishermen expressed dissatisfaction over the massive numbers of ghost jellyfish clogging their nets and the decrease in fish catches. Heavy rainfall (15.82 cm) occurred in and around Chennai during 14-17 October 2024, and subsequently, the bloom of heterotrophic dinoflagellate Noctiluca scintillans occurred along the coastal areas of the Bay of Bengal from Pulicat (North Chennai) to Puducherry on 18-21 October 2024. At night, the blue-green light emerged and the waves were glowing³². The bloom of *B. ovata* is followed by the abundance of scyphozoa jellyfish

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(*C. nozakii*) and hydromedusae (*A. pensilis*). This led to the reduction of copepods, the main predators of diatoms and dinoflagellates, favouring the bloom development of *N. scintillans*.

Though the comb jelly *B. ovata* invaded India through the ships ballast water discharge in the Arabian Sea in 2019, it is now reported in the Bay of Bengal. Since Chennai is the major port area, B. ovata would have been introduced through ballast water discharge. The invasion of alien species causes serious damage to the marine ecosystem. The ctenophore M. leidyi invaded the Black Sea through ship ballast water discharge, resulting in the collapse of pelagic fisheries and an estimated loss of over US\$ 200 million (ref. 33). Anil et al.³⁴ identified the introduction of harmful aquatic organisms and pathogens to new environments via ships' ballast water as one of the four greatest threats to the world's oceans. Jellyfish and ctenophores act as vectors of fish pathogens. There were differences between the bacterial community of B. ovata and the water column^{29,35}. Therefore, a continuous monitoring programme of coastal fishery habitats at the national level is indispensable.

- 1. Afzelius, B. A., The fine structure of the cilia from ctenophore swimming plates. J. Biophys. Biochem. Cytol., 1961, 9, 383–394.
- Franc, J. M., Organization and function of ctenophore colloblasts: an ultrastructural study. *Biol. Bull.*, 1978, 155, 527–541.
- Moroz, L. L., Collins, R. and Paulay, G., Ctenophora: illustrated guide and taxonomy. Chapter 2. In *Ctenophores: Methods and Protocols* (ed. Moroz, L.), Methods in Molecular Biology, 2024, vol. 2757, pp. 27–102; doi:https://doi.org/10.1007/978-1-0716-3642-8_2.
- Raghunathan, C., Mondal, T., Raj, C. P. and Purushothaman, J., Fauna of India Checklist: Ctenophora. Version 1.0. Zoological Survey of India, Kolkata, 2024; https://doi.org/10.26515/Fauna/1/ 2023/Ctenophora.
- Harbison, G. R., Madin, L. P. and Swanberg, N. R., On the natural history and distribution of oceanic ctenophores. *Deep Sea Res.*, 1978, 25, 233–256.
- Tamm, S. L. and Tamm, S., Reversible epithelial adhesion closes the mouth of *Beroe*, a carnivorous marine jelly. *Biol. Bull.*, 1991, 181, 463–473.
- Konsulov, A. S. and Kamburska, L. T., Ecological determination of the new Ctenophora – *Beroe ovata* invasion in the Black Sea. *Oceanol*, 1998, 2, 195–198.
- Shiganova, T. A. and Abyzova, G. A., Revision of Beroidae (Ctenophora) in the southern seas of Europe: systematics and distribution based on genetics and morphology. *Zoo. J. Linn. Soc.*, 2022, 194, 297–322; doi:10.1093/zoolinnean/zlab021.
- https://www.sealifebase.se/Country/CountryList.php?ID=87891&-GenusName=Beroe&SpeciesName=ovata (accessed on 22 November 2024).
- Shiganova, T. A., Bulgakova, Y. V., Volovik, S. P., Mirzoyan, Z. A. and Dudkin, S. I., The new invader *Beroe ovata* Mayer 1912 and its effect on the ecosystem in the northeastern Black Sea. *Hydrobiologia*, 2001, **451**, 187–197.
- 11. Shiganova, T. A., Alekseenko, E. V., Mirzoyan, Z. A. and Kazmin, A. S., Prediction of ctenophore *Beroe ovata* Bruguiere, 1789 habitat expansion and control of invading *Mnemiopsis leidyi* Agassiz, 1865 in the Ponto-Caspian basin associated with climate change. *Mar. Environ. Res.*, 2024, **194**, 106315; https://doi.org/10.1016/j.marenvres.2023.106315.

- Tokarev, Y., Mashukova, O. and Sibirtsova, E., Bioluminescence characteristics changeability of ctenophore *Beroe ovata* Mayer 1912 (Beroida) in ontogenesis. *Turkish J. Fish. Aquat. Sci.*, 2012, 12, 479–484.
- Soto-Angela, J. J. and Burkhardta, P., Reverse development in the ctenophore *Mnemiopsis leidyi*. Proc. Natl. Acad. Sci., 2024, 121(45), e2411499121; https://doi.org/10.1073/pnas.2411499121.
- Vargas, A. M. *et al.*, Morphological and dietary changes encoded in the genome of *Beroe ovata*, a ctenophore-eating ctenophore. *NAR Genom. Bioinform.*, 2024, 6, lqae072; https://doi.org/10.1093/nargab/ lqae072.
- Chacko, P. I., Marine Plankton from waters around the Krusadai Island. Proc. Ind. Acad. Sci., 1950, 31(3), 162–174.
- Rajagopal, P. K., A note on the oxygen uptake of the ctenophore, *Pleurobrachia globosa. Curr. Sci.*, 1963, 32, 319–320.
- Chopra, S., A note on the sudden outburst of ctenophores and medusae in the waters of Bombay. *Curr. Sci.*, 1960, 10, 392–393.
- Goswami, S. C., Occurrence of swarms of Sagitta enflata (Chaetognatha) and Pleurobrachia globosa (Ctenophora) in the coastal waters of Goa. Mahasagar, 1982, 15(3), 163–166.
- Padmavati, G. and FuadSaneen, C. V., Distribution and abundance of gelatinous zooplankton in coastal waters of Port Blair, South Andaman. J. Aquat. Mar. Biol., 2017, 5(6), 175–181; doi:10. 15406/jamb.2017.05.00139.
- Purushothaman, J., Siddique, A., Bhowal, A., Mohan, S., Chandra, K., Benny, N. and Vivek, M. A., Swarms of ctenophore *Pleurobrachia pileus* (O. F. Müller, 1776) in the waters of Sundarban: a menace to the fisheries? *Indian J. Geo Mar. Sci.*, 2020, **49**(06), 1089–1092.
- Praved, P. H., Oliveira, O. M. P., Neethu, K. V., Suhaana, M. N., Bijoy Nandan, S. and Jayachandran, P. R., Unusual blooming of invasive ctenophore *Beroe ovata* (Bruguière, 1789) and geographical range expansion based on morphology and molecular analysis. *Reg. Stud. Mar. Sci.*, 2023, **60**, 102837; https://doi.org/10.1016/j. rsma.2023.102837.
- 22. Shiganova, T. A., Anninsky, B., Finenko, G. A., Kamburska, L., Mutlu, E., Mihneva, V. and Stefanova, K., Black Sea monitoring guidelines. Macroplankton (Gelatinous plankton). EU/UNDP Project: improving of environmental monitoring in the Black Sea – EMBLAS Report, 2021, pp. 1–38.
- Graham, W. M., Pages, F. and Hamner, W. M., A physical context for gelatinous zooplankton aggregations: a review. *Hydrobiologia*, 2001, 451, 199–212.
- Martinussen, M. B. and Bamstedt, U., Digestion rate in relation to temperature of two gelatinous planktonic predators. *Sarsia*, 2001, 86, 21–35.
- 25. Finenko, G. A. *et al.*, Population dynamics, ingestion, growth, and reproduction rates of the invader *Beroe ovata* and its impact on plankton community in Sevastopol Bay, the Black Sea. *J. Plankton Res.*, 2003, **25**(5), 539–549; dx.doi.org/10.1093/plankt/25.5.539.
- Arai, M. N., Predation on pelagic coelenterates: a review. J. Mar. Biol. Ass. UK, 2005, 85, 523–536.
- Pati, S., Food and feeding habits of the chinese pomfret *Pampus chinensis* (Euphrasen, 1788) from the Bay of Bengal. *J. Mar. Biol. Ass. India*, 1977, **19**(1), 44–49.
- Pati, S., Observations on the biology and fishery of black pomfret *Parastromateus niger* (Bloch, 1795) from the Bay of Bengal. *Acta Ichthyol. Piscat.*, 1983, 8(1), 63–74.
- Purcell, J. E. and Arai, M. N., Interactions of pelagic cnidarians and ctenophores with fish: a review. *Hydrobiologia*, 2001, 451, 27– 44.
- Santhakumari, V., Species composition, distribution and abundance of hydromedusae in the exclusive economic zone of the East Coast of India. *Publ. Seto Mar. Biol. Lab.*, 1997, 38, 53–61.
- Shiganova, T. *et al.*, Patterns of invasive ctenophore *Mnemiopsis leidyi* distribution and variability in different recipient environments of the Eurasian seas: a review. *Mar. Environ. Res.*, 2019, 152, 1–23; https://doi.org/10.1016/j.marenvres.2019.104791.

CURRENT SCIENCE, VOL. 128, NO. 3, 10 FEBRUARY 2025

- Tripathi, S. K., Chennai beaches lit up by enchanting bioluminescence: What is it? *India Today*, 2024; https://www.indiatoday.in/ science/story/chennai-beaches-lit-up-by-enchanting-bioluminescencewhat-is-it-2620477-2024-10-21 (accessed on 25 October 2024).
- Knowler, D., Reassessing the costs of biological invasion: *Mnemiopsis leidyi* in the Black Sea. *Ecol. Econ.*, 2005, **52**, 187–199.
- Anil, A. C. *et al.*, Ballast Water Risk Assessment, Ports of Mumbai and Jawaharlal Nehru, India, October 2003: Final Report. GloBallast Monograph Series, International Maritime Organization, London, 2004, vol. 11.
- Daniels, C. and Breitbart, M., Bacterial communities associated with the ctenophores *Mnemiopsis leidyi* and *Beroe ovata*. FEMS. *Microbiol. Ecol.*, 2012, 82, 90–101.

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A report of house rat (*Rattus rattus* L.) damage on phylloclades of dragon plants (*Hylocereus* spp.)

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The dragon plant is an exotic fruit crop gaining importance in urban and peri-urban rooftop cultivation for its nutritional and commercial value. It requires minimal inputs for crop husbandry. Of late an unusual feeding pattern was observed in the phylloclades of dragon plants in the experimental unit of ICAR-Krishi Vigyan Kendra, Kollam. A motion-triggered camera was installed to monitor the pest, which was identified as *Rattus rattus* L. It badly damaged the phylloclades leaving the central core resulting in reduced photosynthetic activity and yield. This clearly highlights the infestation of rats on the phylloclades of dragon plants.

Keywords: Dragon plant, pests, phylloclade damage, *Rattus rattus*, rodent.

THE dragon fruit commonly known as Pitaya or Pitahaya is an important tropical fruit belonging to the family Cactaceae, rich in vitamins, minerals, complex carbohydrates, dietary fibres and antioxidants¹. The genus Hylocereus comprises 18 species². Among them, Hylocereus undatus, Hylocereus polyrhizus and Hylocereus costariscensis are widely cultivated in India spanning an area of 3000-4000 ha (ref. 3). In urban and peri-urban horticulture, it is gaining importance and widely cultivated in terrace or rooftops of Kollam district, Kerala, India (Figure 1). Threats of biotic factors like insects, non-insects and mammalian are commonly found feeding on different fruit crops⁴. There have been reports on feeding damage caused by monkeys, rabbits, squirrels and possums on the lower stems of dragon plants. Similarly, feeding of fruits by monkeys, mice, rats and birds has also been reported⁵. However, the present study reports the feeding damage of dragon stem (phylloclades) by rats.

The present study was conducted at the demonstration unit of dragon plants maintained on the terrace of ICAR-Krishi Vigyan Kendra (KVK), Kollam, during October-November 2024. The climate is tropical, having a lat. 8.987°N and long. 76.82°E. The plants in the demonstration unit showed severe feeding damage on the phylloclade without the physical presence of any kind of insect pests even after continuous monitoring during daytime. This prompted further probe on this. The dragon plants were monitored for the pest activity using a HI XECURE, 4G, solar powered, motion-triggered CCTV camera with dual lens connected to an internet connectivity mounted on a wall at a height of 1.4 m from the ground enabling a bird's-eye view, spanning over the experimental plants round the clock. A second camera was fixed to monitor the ground activity of the suspected pest. The visuals were recorded and monitored continuously for a week to confirm the nature of damage, frequency and pattern of the visit of the pest. Camera visuals revealed the presence and feeding activity of rodents in the experimental unit. To confirm and identify the species of the rodent, a live trap was installed in the experimental unit and monitored regularly. Additionally, an extensive survey was conducted among the urban and periurban dragon fruit growers, for similar infestation and feeding symptom, with the support of several line departments including Agricultural Technology Management Agency (ATMA), State Horticulture Mission of Department of Agriculture and Farmers Welfare, Government of

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