

## Sulphate Reducing Bacteria in South-West Coastal Aquaculture System of India

A. NINAWÉ\* and S. BANIK \*

Central Marine Fisheries Research Institute,

P. O. Box-2704, Cochin-682031

Ecological aspects viz, temperature, pH, Eh salinity and concentration of dissolved oxygen, hydrogen sulfide and sulfate reducing bacteria in brackish water environments of two different culture practices were studied in summer and after onset of monsoon to clarify the major factors governing their distribution, seasonal fluctuation and their rate of  $H_2S$  production. A comparative study regarding distribution of sulfate reducing bacteria and  $H_2S$  production from both the culture systems has been discussed. Ten selected isolates of sulfate reducing bacteria from sediments were identified as species of the genus *Desulfovibrio*.

South West coastal aquaculture of India is a unique system to cultivate prawn in paddy fields after harvesting. Due to high salinity of the soils, cultivation of salt tolerant variety is the practice. But occasional fish mortality has been recorded when water of such culture ponds turned black. This blackening of water and sediment at frequent intervals has been thought as activity of heterotrophic sulfate reducing bacteria (Unnithan, 1985). Sulfate reducing bacteria grow in saline to nonsaline environments (Abd-el Malek and Rizk, 1963) favourably at near neutral pH when redox potential reaches as low as -100 mV (Nedwell, 1982) and at mesophilic range of temperature (Kimata et al., 1955), although aerobic forms has been found (Jørgensen, 1977). Ecological aspects of sulfate reduction in nature, distribution of sulfate reducing bacteria and their metabolic activity in situ in relation to organic matter content of their habitat and concentration of sulfate ions varies widely (Tezuka, 1979).

So, study was undertaken to search active sulfate reducing bacteria in prawn and prawn cum paddy culture fields and to evaluate their role in producing  $H_2S$  in vitro which causes fish mortality, beyond toxic level (Adelman and Smith, 1970; Smith and Oseid, 1974).

### MATERIALS AND METHODS

#### *Habitat*

The study was conducted in the brackish water experimental ponds in

---

\*A. Ninawe, Department of Science and Technology, New Delhi, India.

\*\*Dr S. Banik, Jute Technological Research Laboratories, 12 Regent Park, Calcutta-700 040, India.



Narakkal region of Vypeen island, Cochin, Kerala, India. The ponds are connected to the main canal through a sluice gate where the exchange of water through out the period of study was influenced by tides, and artificially prawn seeds were stocked (Muthu, 1978). During the period of October to May prawn cum paddy culture fields are utilized for cultivation of prawn followed by cultivation of a salt tolerant variety of paddy, viz., 'Pokkali' from June to September whereas, the perennial ponds are used to cultivate prawns only being harvested twice a year.

### *Sampling*

Samples were collected in the month of February—summer and May—two weeks after onset of monsoon in the year of 1984. Undisturbed mud cores were collected from three sites for each sampling by a specially designed hand corer upto a depth of 5 cm, homogenised and composited and was stored in a sterilized glass bottle. Water samples were collected in sterilized glass bottles for bacteriological analysis. Samples were collected in glass stoppered bottles without allowing any air bubble and fixed immediately with Winkler A and B solutions for determining dissolved oxygen content chemically. Winkler A is a 20%  $\text{MnCl}_2$  solution and Winkler B is 25% KI in 41% NaOH in water.

### *Experimentals*

Both atmospheric as well as water temperature were recorded using a Hg thermometer while collecting the samples. pH and Eh of the collected samples were determined immediately (Cohen 1957). Bacteriological analyses were carried out on the other chemical parameters of sediment and water were determined on the same day of collection. Salinity of the water samples were estimated following Mohr—Knudson argentometry and dissolved oxygen (D.O.) in water was determined according to modified Winkler's method (Strickland and Parsons, 1968). The pH value of the water and sediment samples were determined by an Elico pH meter, using glass electrode. Eh of the water and sediment samples were found out by the same pH meter using a Pt electrode. Total sulfate content in water and sediment and hydrogen sulfide content in water were analysed by using turbidimetric and titrimetric methods respectively (A.P.H.A., 1971).

Sulfate reducing bacteria from both water and sediment were enumerated using Postgate's medium (Starr et al., 1981) modified by the use of 50% (V/V) aged sea water as a dilutant and 2% (W/V) agar following pour plate technique. Sodium thioglycollate was added to ensure anaerobiosis.



and the pH was adjusted to 7.4. Plates were incubated anaerobically in jar containing nitrogen and carbon dioxide gas for seven days and only black coloured colonies were counted for enumerating sulfate reducing bacteria (S.R.B.). Hydrogen sulfide production rate was studied in vitro by providing 200 ml of Postgate's nutrient broth (Starr et al., 1981) containing thioglycollate in 500 ml round bottom flasks. After inoculation of 1.0 g fresh sediment, flasks were incubated for 10 days at  $30 \pm 1^\circ\text{C}$ . Anaerobic condition was maintained throughout the incubation period by adding a layer of sterilized liquid paraffin. The released  $\text{H}_2\text{S}$  gas accumulated in tubes by replacing water due to reduction of sulfate was noted. Bacterial isolates were taken from sediments only and maintained in the same liquid medium (Starr et al., 1981) in a screw cap bottle covered with sterilized liquid paraffin as above. Characterization of the isolated bacteria were carried out with the help of taxonomic scheme given in Bergey's manual of determinative bacteriology (Buchanan and Gibbons, 1974).

## RESULTS AND DISCUSSIONS

From the data presented in Table 1 it is indicated that atmospheric and water temperature did not vary significantly due to seasonal change. pH values of water raised with the onset of monsoon but the same effect in sediment was found in prawn cum paddy cultivation fields only. Eh values of sediments were far more reduced than that of water irrespective of season and no definite seasonal effect was noticed. Salinity of water and sulfate content in both water and sediments were declined in rainy season. Dissolved oxygen content and  $\text{H}_2\text{S}$  concentration in water was found to increase during those period in comparison to summer.

Rise of pH values of water in monsoon may be due to increased ionization of solutes as a result of dilution effect and poor buffering capacity but the case of sediment was not understood and needs further study. Higher anaerobic condition in sediments might be due to total removal of oxygen as a result of aerobic microbial activity. Decrease of salinity in water as well as decrease of sulfate ion content in both water and sediment was probably due to dilution effect. However, reverse trend in dissolved oxygen content in water might be due to addition of oxygen saturated downpour and more turbulence in water (Ramamirtham, 1968). Higher accumulation of  $\text{H}_2\text{S}$  in water might be consequence of reduction of sulfate in both water and sediment.

From data presented in Table 2, a declining trend of sulfate reducing bacteria was observed from summer to monsoon and the decrease of sulfate

Table—1 Ecological aspects of the environment

| Date             | Pond | Atmospheric Water   |                     | Atm.                             |               |         |               | Water analysis |                            |                     |                            | Sedi-<br>ment<br>analy-<br>sis<br>SO <sub>4</sub> <sup>-2</sup><br>(mg/g) |   |
|------------------|------|---------------------|---------------------|----------------------------------|---------------|---------|---------------|----------------|----------------------------|---------------------|----------------------------|---|---|
|                  |      | temperature<br>(°C) | temperature<br>(°C) | PH                               |               | Eh (mV) |               | (mm of Hg)     | Sali-<br>nity mg/<br>(ppt) | D.O.<br>lit. mg/lit | H <sub>2</sub> S<br>mg/lit |   | SO <sub>4</sub> <sup>-2</sup><br>mg/lit |
|                  |      |                     |                     | Water                            | Sedi-<br>ment | Water   | Sedi-<br>ment |                |                            |                     |                            |   |   |
| 24.2.84          | I    | 31.5                | 32.0                | 7.95                             | 7.90          | +145    | -65           | 769.14         | 17.58                      | 3.95                | 0.018                      | 28.0  | 11.5                                    |
|                  | II   | 31.5                | 32.0                | 7.45                             | 7.15          | +135    | -120          |                | 17.54                      | 3.18                | 0.018                      | 24.0  | 17.0                                    |
| 24.5.84          | I    | 33.5                | 34.5                | 8.40                             | 7.80          | +70     | -155          | 760.94         | 16.24                      | 4.83                | 0.546                      | 18.0  | 9.8                                     |
|                  | II   | 33.5                | 34.0                | 8.40                             | 7.60          | +65     | -100          |                | 15.23                      | 4.66                | 0.617                      | 16.4  | 13.7                                    |
| I—Perennial pond |      |                     |                     | II—Prawn-cum-paddy culture field |               |         |               |                |                            |                     |                            |   |   |

Table—2 Enumeration of sulfate reducing bacteria and sulfate reducing power of sediments

| Date             | Pond | Sulfa reducing bacteria |                      | Ratio of sulfate reducing bacteria in water and sediment | Before inoculation in medium     |     | After incubation in medium |      | mg H <sub>4</sub> S released from 200 ml Postgate's broth |
|------------------|------|-------------------------|----------------------|--|----------------------------------|-----|----------------------------|------|---|
|                  |      | Water per ml.           | Sediment per g.      |  | PH                               | Eh  | PH                         | Eh   |   |
| 24.2.84          | I    | 17 × 10 <sup>3</sup>    | 21 × 10 <sup>4</sup> | 1 : 12.35  | 7.65                             | —45 | 7.0                        | —245 | 27.8  |
|                  | II   | 25 × 10 <sup>3</sup>    | 28 × 10 <sup>4</sup> | 1 : 11.20  |                                  |     | 6.5                        | —270 | 53.2  |
| 24.5.84          | I    | 5 × 10 <sup>3</sup>     | 18 × 10 <sup>3</sup> | 1 : 3.60   | 7.50                             | —75 | 6.8                        | —235 | 38.8  |
|                  | II   | 6.6 × 10 <sup>3</sup>   | 25 × 10 <sup>3</sup> | 1 : 3.79   |                                  |     | 6.5                        | —265 | 47.6  |
| I—Perennial pond |      |                         |                      |  | II—Prawn-cum-paddy culture field |     |                            |      |   |



Table—3 Differential biochemical characteristics of the isolated bacteria

| Number of isolates     | D <sub>1</sub> | D <sup>2</sup> | D <sup>3</sup> | D <sup>4</sup> | D <sup>5</sup> | D <sup>6</sup> | D <sup>7</sup> | D <sup>8</sup> | D <sup>9</sup> | D <sup>10</sup> |
|------------------------|----------------|----------------|----------------|----------------|----------------|----------------|----------------|----------------|----------------|-----------------|
| Growth in :            |                |                |                |                |                |                |                |                |                |                 |
| Pyruvate plus sulfate  | +              | +              | +              | +              | +              | +              | +              | +              | +              | +               |
| Pyruvate minus sulfate | +              | +              | +              | +              | +              | +              | +              | —              | +              | +               |
| Malate plus sulfate    | +              | +              | +              | +              | —              | +              | —              | +              | +              | +               |
| Malate minus sulfate   | —              | —              | —              | —              | —              | —              | —              | —              | —              | —               |
| Choline plus sulfate   | +              | +              | +              | +              | +              | +              | +              | —              | +              | +               |
| Choline minus sulfate  | +              | +              | +              | +              | +              | +              | +              | —              | +              | +               |
| Salt requirement       | 3%             | 3%             | 3%             | 3%             | 3%             | 3%             | 3%             | 3%             | 3%             | 3%              |
| Identified as          | 1              | 1              | 1              | 1              | 2              | 1              |                | 2              | 1              | 1               |

+ Reaction positive  
 — Reaction negative

- 1 *Desulforibrio desulfuricans* sub. *aesturii*
- 2 *Desulfovibrio vulgaris* sub. *oxamicus*
- 3 *Desulfovibrio salexigens*

reducing bacteria in sediment was more than that of water. The number of SRB in prawn cum paddy culture field was always higher in comparison to perennial ponds. pH values in Postgate medium reached near to neutrality whereas the respective  $E_h$  values reduced considerably as a result of growth of SRB. Higher rate of sulfate reduction was noticed, as evidenced by release of  $H_2S$ , in prawn cum paddy culture field samples in vitro, in compared to perennial ponds. In perennial ponds rate of sulfate reduction was increased in rainy season as compared to summer but the reverse observation was found in prawn cum paddy cultivation fields.

The decline of sulfate reducers in monsoon may be due to decreased salinity (Kimata et al., 1955), reduced concentration of sulfate (Howarth and Teal, 1979), increased concentration of oxygen and increased pH (Nedwell, 1982). Relatively more decreased of sediment bacteria might be due to their greater sensitivity towards environmental changes. Higher accumulation of SRB in prawn cum paddy culture fields might be due to availability of organic matter in higher concentration (Nedwell, 1982), and other essential mineral nutrients added through decomposition of plant remains. Recorded pH and  $E_h$  values in vitro experiment indicate the choice of sulfate reducers of the habitat similar as described by Zo Bell (1958). Higher rate of sulfate reduction in prawn cum paddy cultivation fields as compared to perennial prawn culture ponds might be due to thriving higher number SRB in those habitat.

Table 3 indicates that all the isolated sulfate reducing bacteria from sediment were members of the genus *Desulfovibrio*. All bacteria were Gram negative, asporogenous rods, producing black coloured round colonies in medium containing pyruvate in presence of sulfate and have optimum salt requirement of 3%. Seven of the isolates were identified as *Desulfovibrio desulfuricans* sub sp. *aesturii*, two of them as *Desulfovibrio vulgaris* sub sp. *oxamicus* and one as *Desulfovibrio selexigens*.

From the present study it was revealed that among heterotrophic sulfate reducing bacteria, *Desulfovibrio desulfuricans*, *Desulfovibrio selexigens* and *Desulfovibrio vulgaris* were active in sediments of the habitat and they can produce significant amount of  $H_2S$  in nature to produce toxicity when condition favours.

#### ACKNOWLEDGEMENTS

The authors are grateful to the Director, C. M. F. R. 1, Cochin for the facilities provided for conducting the research work.



## REFERENCES

- Abd-el-Malek, Y. and Rizk, S. G. (1963). Bacterial sulfate reduction and the development of alkalinity. III. Experiments under natural conditions. *J. Appl. Bacteriol.* **26**, 20-26.
- Adelman, I. R. and Smith, L. L. (1970) Effect of hydrogen sulfide on northern pike eggs and sac. fry. *Trans. Amer. Fish. Soc.* **99**, 501-509.
- American Public Health Association. (1971). Standard methods for the examination of water and waste water. 13th edn. Washington. 974.
- Buchanan, R.E. and Gibbons, N. E. (eds) (1974). Bergey's manual of determinative bacteriology. 8th edn. Williams and Wilkins.
- Cohen, B. (1957). The measurement of pH, titrable acidity and oxidation reduction potentials. In: *Manuals of Microbiological Methods*. McGraw Hill Book Co. New York. 72-98.
- Howarth, R. W. and Teal, J. M. (1979) Sulfate reduction in a New England salt marsh. *Limnology and Oceanography*. **24**, 999-1013.
- Jørgensen, B. B. (1977). Bacterial sulfate reduction within reduced microniches of oxidized marine sediments. *Marine Biology*, **41**, 7-17.
- Kimata, M., Kobota, H., Hata, Y., and Tajima, T. (1955). Studies on the marine sulfate reducing bacteria. II. Influences of various environmental factors upon the sulfate reducing activity of marine sulfate reducing bacteria. *Bull. Jap. Soc. Sci. Fish.* **21**, 109-112.
- Muthu, M. S. (1978). A general review of penaeid prawn culture. In: C. M. F. R. I. special publication **3**. 73-106.
- Nedwell, D. B. (1982). The cycling of sulphur in marine and fresh water sediments. In: Nedwell, D. B. and Brown, C. M. (eds) *Sediment Microbiology* Academic Press. 73-106.
- Ramamirtham, C. P. (1968). Vertical distribution of temperature, salinity and dissolved oxygen in the Maldivian region of the Indian Ocean. *Indian J. Fish.* **15**, 27-29.
- Smith, L. L. and Osleid, D. M. (1974). Effect of hydrogen sulfide on development and survival of eight fresh water fish species. *Int. Symp. on early life history of fish*. Oban, Scotland 17-23.
- Starr, M. P., Stolp, H., Truper, H. G., Bblows, A., and Schlegel, H. G. (eds). (1981). In: *The prokaryotes. A hand book on habitats, isolation and identification of bacteria*. Vol. 1. Springer Verlag.
- Strickland J. D. H., and Parsons, T. R. (1972) A practical handbook of sea water analysis. Fisheries Research Board of Canada, Ottawa.
- Tezuka, Y. (1979). Distribution of sulfate reducing bacteria and sulfides in aquatic sediments. *Jap. J. Ecol.* **29**, 59-102.
- Unnikrishnan, K. A. (1985). A guide to prawn farming in Kerala. C. M. F. R. I. special publication No. 21. Cochin, India.
- Zobell, C. E. (1953). Ecology of sulfate reducing bacteria. *Producers Monthly*. **22**, 12-29.