Screening of seaweed extracts against antibiotic resistant post operative infectious pathogens

S. RAVIKUMAR, L. ANBURAJAN, G. RAMANATHAN AND N. KALIAPERUMAL¹

Department of Marine Biotechnology, Institute for Coastal Area Studies Manonmaniam Sundaranar University, Scott Christian College Campus Nagercoil - 629 003, India ¹Regional Centre of Central Marine Fisheries Research Institute Mandapam Camp - 623 520, India

ABSTRACT

Fifty five seaweed extracts belonging to 11 species of seaweeds were tested against post operative infectious drug resistant bacteria viz., E. coli, Klebsiella pneumoniae, Pseudomonas aeruginosa, Streptococcus pyogens, Staphylococcus aureus. Among the seaweed extracts, the acetone extracts of Caulerpa cupressoides shows maximum inhibitory activity against E. coli and propanol extracts of Gracilaria edulis shows maximum inhibitory effect against K. pneumoniae. Acetone extracts of Padina tetrastromatica and Laurencia cruciata show maximum inhibitory activity against P. aeruginosa, butanol extracts of Hypnea musciformis, Caulerpa cupressoides and Chaetomorpha linoides show maximum inhibitory effect against S. aureus.

Introduction

Hospital associated diseases are called nosocomial infections. It develops in the patient during stay in the hospital. A recent study indicates that 5 to 19% of the patients in acute care hospitals develop a nosocomial disease. Bacteria are the leading cause of nosocomial disease and viruses are a distant second. Occasionally fungi cause disease but rarely protozoa are involed. Mostly nosocomial diseases are caused by gram negative bacilli like *E. coli*. The use of therapeutic and diagnostic equipment (such as intravenous and urinary catheters), surgical procedure and transplantation has increased the risk of

nosocomial diseases. Increased antibiotic therapy leads to the development of drug resistance against wide spectrum of antibiotics. Hence, this study has been undertaken to find out the new drugs from marine living resources.

Multiple drug resistance organisms are becoming common causes for infections in acute and long term care units in hospitals (Emori and Gaynes, 1993). The emergence of the resistant bacteria has created a major concern and an urgent need for new antibacterial agents (American society of Microbiology, 1995; Davis 1994; Spratt, 1994). Marine environment is an exceptional reservoir of biologically active natural products, many

of which exhibit structural features not been found in terrestrial natural products (Ireland et. al., 1988). Marine natural products, the secondary or non-primary metabolites produced by living organisms, have been exploited by people for a variety of purposes including their use as food, fragrance, pigments, insecticides and medicines. Each year, an increasing number of novel marine metabolites are reported indicating that the marine environment is likely to continue to be a prolific source of new natural products, for many years to come. Approximately 2500 new metabolites were reported from a variety of marine organisms during the decade from 1977-1987 (Ireland et. al., 1993). Because of the diversity of marine organisms in various habitats, marine natural products encompass a wide variety of chemical classes including terpenes, shikimates, polyketides, acetogenins, peptides and alkaloids of varying structures and multitude of compounds of mixed biosynthesis.

Materials and Methods Collection of seaweed samples

Fresh seaweed samples of Caulerpa cupressoides, Chaetomorpha linoides, Enteromorpha intestinalis, Gracilaria edulis, Colpomenia sinuosa, Hypnea musciformis, Laurencia cruciata, Padina boergesenii, P. tetrastromatica, Sargassum wightii and Ulva lactuca were collected from the south west coast of India. Healthy and well grown plants were collected and cleaned with seawater and then freshwater to remove all epiphytes etc. and dried in room temperature. The dried samples were powdered for the extraction of antimicrobial compounds.

Isolation of drug resistant POI pathogens

Pus samples were collected from 5 male and 5 female patients from post operative wounds by touching the infected area with a sterile swab (antiseptic agent was not applied

before taking the specimen). The collected swabs were transferred immediately in a sterile test tube and then into a transport medium. Two swabs were collected of which one was used for direct examination and another swab was inoculated into the solid media (Blood agar and Mac conkey's agar) and incubated at 35°C for 18-24 hrs. After incubation, the morphology of the colonies were observed and identified as *E. coli, K. pneumoniae, P. aeruginosa, S. pyogens, S. aureus* based on Bergey's manual of determinative bacteriology (Holt *et. al.*, 1994). Identified strains were checked for antibiotic resistance by following the method of Bauer *et. al.* (1996).

Extraction and bioassay of antimicrobial compounds

Dried powder of (50 g) seaweed samples were immersed in various solvents (Acetone, n - Butanol, propanol, water and Benzene). Percolation were carried out three times and kept for evaporation. The dried extract powder was redissolved in the respective solvents and used for the bioassay. Twenty milligram of evaporated extracts were impregnated into a sterile filter paper disc (5 mm diameter). The impregnated extracts and commercial antibiotic discs (30 antibiotics) were placed on seeded agar plates and kept it for incubation. After 24 hrs of incubation, the zone of inhibition of the bacterial species were measured by using graduated scale and expressed as millimeter in diameter.

Results and Discussion

There are five bacterial species isolated from post operative infectious wounds and identified as *E. coli*, *K. pneumoniae*, *P. aeruginosa*, *S. pyogens*, *S. aureus*. These strains were screened for antibiotic resistance with 30 antibiotics. The resistant antibiotics against each bacterial strain are presented in Table 1. Eleven crude seaweed extracts were

Table 1. List of antibiotics resistant to the five bacterial pathogens

Name of the Antibiotics	Name of the bacterial pathogens							
Antibiotics	E.c	K.p	P.a	S.p	S.a			
Ceftazidine	R	R	R	R	R			
Lincomycin	R	R	R	R	S			
Vancomycin	R	S	R	S	S			
Co-Trimazine	R	R	R	R	R			
Nitrofurantoin	R	R	R	R	S			
Streptomycin	R	R	S	S	S			
Cephalothin	R	R	R	R	S			
Clindamycin	R	S	R	R	R			
Cloxacillin	R	R	S	R	R			
Oxytetracycline	R	R	R	S	S			
Penicillin - G	R	R	R	S	S			
Co-Trimoxazole	R	R	R	S	S			
Erythromycin	R	S	R	R	R			
Amoxycillin	R	S	S	S	S			
Cephalexin	R	R	S	S	S			
Cephaloridine	R	S	S	S	S			
Methicillin	R	S	S	S	S			
Oleandomycin	R	S	S	S	S			
Nallidixic acid	R	S	S	S	S			
Ampicillin	R	R	R	S	S			
Colistin	R	S	S	S	S			
Oxacillin	R	S	S	S	S			
Bacitracin	R	S	R	S	S			
Polymycin	R	S	S	S	S			
Neomycin	R	S	S	R	S			
Cephalotoxine	S	R	S	S	S			
Carbenicillin	S	R	S	S	S			
Gentamycin	S	R	S	S	S			
Norfloxacin	S	R	S	S	S			
Furazolidone	S	S	S	R	S			

E.c - Escherichia coli, K.p - Klebsiella pneumoniae, P.a -Pseudomonas aeruginosa, S.p - Streptococcus pyogens, S.a - Staphylococcus aureus tested against drug resistant post operative infectious pathogens (Table 2). Among the seaweed extracts, the acetone extract of Caulerpa cupressoides gives maximum inhibitory activity (9 mm) against the bacterium E. coli. The propanol extract of Gracilaria edulis gives maximum inhibitory activity (9 mm) against the bacterial pathogen K. pneumoniae. The acetone extract of Padina tetrastromatica and Laurencia cruciata gives maximum inhibitory activity (10 mm) against P. aeruginosa. The butanol extract of Hypnea musciformis, Caulerpa cupressoides and Chaetomorpha linoides gives the maximum inhibitory activity (8 mm) against S. pyogens. The butanol extract of Padina tetrastromatica gives the maximum inhibitory activity (10 mm) against S. aureus. Among the five species of pathogens, the inhibitory activity was in the series of Pseudomonas aeruginosa, S. aureus, E. coli, K. pneumoniae and S. pyogens.

The present study reveals the poor activity of water extract in all the test plant samples. Among the organic solvent extracts, the butanol, propanol and acetone extracts and not of benzene were highly effective against all post operative pathogens. Best and higher activity type was found in Padina tetrastromatica and Laurencia cruciata against Pseudomonas aeruginosa followed by Padina boergesenii extract against S. aureus. Pesando and Caram (1984) reported that E. coli was more sensitive to Laurencia obtusa. Issac and Hedge (1987) found that E. coli was more sensitive to Sargassum johnstonii. Glombitza (1970) suggests that dimethyl sulphide and acrylic acid from seaweeds has antimicrobial activity. Pratt et. al. (1951), observed that Spirogyra sp. was known to reduce typhoid,

R - Resistance: S - Sensitive

Table 2. Sensitivity pattern of seaweed extract against five species of multiple drug resistant nosocomial infectious pathogenic bacteria.

Name of the seaweed	Solvent	Zone of inhibition in mm			on	Name of the seaweed	Solvent	Zone of inhibition in mm					
		E.c	K.p	P.a	S.p	S.a			E.c	K.p	P.a	S.p	S.a
Padina tetrastromatica	Benzene	NS	NS	6	6	6		Benzene	7	6	6	NS	8
	Butanol	7	6	NS	NS	9		Butanol	7	8	8	8	6
	Propanol	8	8	7	6	8	Chaetomorpho	a Propanol	6	8	6	6	NS
	Acetone	NS	NS	10	7	6	linoides	Acetone	NS	NS	5	6	NS
	Water	6	NS	NS	NS	NS			1 10			-	
	Benzene	6	6	6	7	7		Water	6	NS	NS	NS	NS
7.7	Butanol	8	5	6	8	8		Benzene	6	6	6	6	6
Hypnea	Propanol	7	6	6	7	6		Butanol	6	7	NS	NS	8
musciformis	Illya	Ulva	Propanol	6	6	6	NS	7					
Ben	Water	6 6	NS 7	NS 5	10 5	NS 5	lactuca	Acetone	NS	NS	8	NS	NS
	Benzene Butanol	7	5	5 7	NS	5 6		Water	NS	NS	NS	NS	NS
Laurencia	Propanol	6	NS	NS	6	5				NS			
cruciata	Acetone	8	NS	10	7	NS		Benzene	6		6	6	6
	Water	6	7	NS	NS	NS		Butanol	8	8	7	NS	9
	Benzene	6	6	5	6	6	Sargassum	Propanol	6	6	6	6	5
	Butanol 7 NS 7 NS 6	wightii	Acetone	NS	NS	NS	NS	NS					
Caulerpa	Propanol	7	NS	8	7	6		Water	NS	NS	NS	NS	NS
cupressoides	Acetone	9	6	5	8	7		Benzene	6	6	6	6	6
*	Water	6	6	NS	NS	NS		Butanol	8	8	7	7	7
	Benzene	6	NS	6	6	6	Gracilaria edulis		-	-			
	Butanol	7	7	6	7	6		Propanol	5	9	6	7	6
Enteromorpha	Propanol	6	NS	7	7	7	eautis	Acetone	NS	NS	5	6	5
intestinalis	Acetone	NS	NS	NS	NS	NS		Water	NS	NS	NS	NS	NS
5. 11	Water	6	10	NS	NS	NS		Benzene	7	6	6	NS	NS
	Benzene	6	6	6	NS	NS		Butanol	8	8	8	7	6
	Butanol	8	7	NS	NS	10	Colpomenia		6	NS	7	7	6
Padina	Propanol	6	6	7	7	7	sinuosa	ropunor	-			,	-
	Acetone	7	NS	5	NS	NS	3	Acetone	NS	NS	5	NS	NS
	Water	5	NS	NS	NS	NS		Water	NS	NS	NS	NS	NS

E.c - Escherichia coli, K.p - Klebsiella pneumoniae, P.a Pseudomonas aeruginosa, S.p - Steptococcus pyogens, S.a - Staphylococcus aureus NS - No Sensitivity

cholera, dysentry. Padmini Sreenivasa Rao and Karmarkar (1986) reported that ethanol extract of *S. johnstonii, Enteromorpha* sp and *Corallina officinalis* showed high activity against *S. aureus* than the extracts of acetone and n - butanol. Xixu and Lee (2001) reported that the plant flavanoids highly inhibited the growth of multiple drug resistant *Staphylococcus aureus* and *Burkholderia cepacia*.

To our knowledge this is the first report from India of bioactive compounds effective against the multiple drug resistant bacteria.

Acknowledgements

The authors are grateful to the authorities of M. S. University, Tirunelveli for providing facilities and to the Tamilnadu State Council for Science and Technology for sanctioning the Student Project.

Literature cited

- Bauer, A. W., W.W.M. Kirby and J. C. Sherris 1996. Antibiotic susceptibility testing by a single disc method. *Am. J. Clin. Pathol.*, 45:493.
- Davis, J., 1994. Inactivation of antibiotics and the dissemination of resistance genes. *Science*, 264:375-381.
- Emori, T. C. and R. Gaynes 1993. An overview of nosocomial infections, including the role of the microbiology laboratory. *Clin. Microbiol. Rev.*, 6:428-442.
- Glombitza, K.W. 1970. Antimicrobial constituents in algae. Quantitative determination of acrylic acid in marine algae. *Planta medica*, 18:210-221.
- Holt, J. G., N.R. Krieg, P.H. Sneath, J.T. Staley and T. Williams 1994. *Bergey's manual of determinitive bacteriology 9th edn*, William & Wilkins. Baltimore. 260 pp.
- Ireland, C.M., D.M.Roll, T.F.Molinski, T.C.Mckee, T.M. Zarbriske and J.C. Swersey 1988. Uniqueness of the marine environment: categories of marine natural products from invertebrates In: *Biomedical importance of marine organisms, California Academy of Scince, California*. 41 47 pp.

- Ireland, C., B. Copp, M. Foster, L. Mc-Donald, D. Rdisky and J. Swersey 1993. Biomedical potential of marine natural products. *Mar. Biotechnol.*, 11:1-43.
- Isacc, W., and G. R. Hedge 1987. Fresh water algae of Karnataka state Certain taxa from Uttarakannada District, new to the flora of Karnataka. *Phykos*, 26: 123 129.
- Padmini Sreenivasa Rao, P. and S. M. Karmarkar 1986. Biological investigation of genus Sargassum - antifungal activity of fractions of different species of Sargassum. Phykos, 25:6-11.
- Pesando, D. and B. Caram 1984. Screening of marine algae from the French mediterranean coast for antibacterial and antifungal activity. *Bot. Mar.*, 27:381-386.
- Pratt, R., R. Maunter, G. M. Gardner, V. Sha and J. Dutrendy 1951. Report on antibiotic activity of seaweed extracts. *J. Amer. Pharm. Asst. Sci.*, 40:575-579.
- Spratt, B. G. 1994. Resistance to antibiotics mediated by target alterations. *Science*, 264: 388-393.
- Xixu, H., and F. Lee 2001. Activity of plant flavanoids against antibiotic resistant bacteria. *Phytother. Res.*, 15:39-43.