## Static bioassay with *Liza parsia* exposed to DDVP, an organophosphate

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DDVP, a water-soluble organophosphate insecticide, is widely used in the Kolleru region of Andhra Pradesh for the control of ectoparasites, such as Lernea and Argulus, on fishes (Muthu el al. 1988). DDVP is used as medicine in salmon farming against the sea lice. The use of this chemical in salmon fanning, however, appears to have deleterious effects on marine invertebrates (Egidius and Moester 1987). Stephanie Pain (1989) linked the epidemic of eye disease in salmon of the wild to the use of this chemical in farms. Acute toxicity tests of DDVP were, therefore, conducted on Liza parsia. Among mullets, Liza parsia, along with Mugil cephalus, has gained considerable importance in fish culture because of its resistance to environmental changes and easy availability. It was, therefore, selected for this study.

*Uia parsia* of 85—120 mm size and 6.50—13.25 g weight were collected live by cast nets from brackishwater canals of Puduvypeen area in Cochin. They were acclimatized to laboratory condition for about 2 weeks by maintaining in salinity  $10.0 \pm 1.0\%$ , pH 6.0±0.5 and temperature  $27.5^{\circ}\pm 1.5^{\circ}$ C. To avoid fungal attack the medium was treated with 11 mg of malachite green per 100 litres of water. The fish were fed

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once a day.

The commercial grade DDVP composed of Dichlorvos 76% m/m, emulsifier 10.6% m/m and solvent 13.4% m/m was used for the preparation of stock solution.

A range-finding bioassay was conducted as per APHA-AWWA-WPCF (1975) and Reish and Oshida (1987). No mortality was observed at 0.1 ppm but 75% mortality occurred at 1 ppm at 96 hr. Hence the concentrations between 0.1 and 1.2 ppm were selected for bioassay procedure.

Static bioassay method (Reish and Oshida 1987) was used in the entire experiment. Each bioassay consisting of a series of 6 test concentrations and a control was used. Each concentration was run in duplicate

Table 1. Mortality at different expoiure periodi obtained in *Lisa parsia* at different concentration! of DDVP\*

Concen-	Animali	Mortality (No.) after					
tration (ppm)	releaied (No.)	24 hr	48 hr	72 hr	96 hr		
0.2	16	0	2	2	2		
0.4	16	2	4	5	6		
0.6	16	4	5	7	9		
0.8	16	6	9	11	13		
1.0	16	7	10	13	16		
12	16	10	11	14	16		

\*Compo\*ition of DDVP was: Dlchlorvoi 76% mAn, emubifier 10.6% m/m and solvent 13.4% mAn.



Fig.l .LC<sub>M</sub> values of DDVP for *Liza parsia* after 24,48,72 and 96 hr of exposure. A. Response curve for 24 hr leihal concentration (LC<sub>n</sub> « Anti log (0.006) = 1.015 ppm). B. Response curve for 48 hr lethal concentration (LC<sub>n</sub> • Anti log (-0.125) • 0.750 ppm). C. Response curve for 72 hr leihal concentration (LC<sub>M</sub>=Ami log (-0.256)=0.554 ppm). D. Response curve for 96 hr lethal concentration (LC<sub>W</sub> = Anti log (-0.317) = 0.482 ppm).

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(APHA-AWWA-WPCF 1975). Precautions were taken to avoid contamination of the controls. Test animals were not fed during the experiment. The percentage of survival at die end of every 24, 48, 72 and 96 hr was accounted (Table 1). The data were processed by Probit analysis (Reish and Oshida 1987) in computer. The LC<sub>M</sub> values obtained for 24,48,72 and 96 hr were 1.014725, 0.750109, 0.554255 and 0.482347 respectively. The percentage mortality vs log concentrations were plotted in probability papers (Fig. 1) and the LC<sub>M</sub> values were got graphically (Seegert et al. 1979) also (Table 2). The slope function, 95% confidence limit and 95% fiducial limits (upper and lower) were calculated using the formulae from response curves for different exposure times after Reish and Oshida (1987).

Slope (S)

95\* confidence limit JVTT

where N, total number of organisms tested at those exposure concentrations whose expected results were between 16% and 84%; and 2.77, a constant.

95% fiducial limits are :

Upper limit =  $LC_M X LC_{SO}$ 

Lower limit:  $tic_{,50}^{so}$ 





Fig. 2. Lethal concentration v e n a time curvet.

The LCjgValues showed gradual decrease with increase in time and 95% fiducial limits of each of response curves for different exposure periods showed decreasing trend (Table 2). The 95% confidence limit and slope function shown against each LC<sub>M</sub> values have overall decreasing trend except for a slight increase in 48 hr LC<sub>M</sub>.

The LC<sub>M</sub> LCJI and LC<sub>1(</sub> obtained from response curves for 24, 48, 72 and 76 hr (Table 2) were graphically plotted for non lethal concentrations (Fig. 2) which showed that for a short application a concentration around  $\theta 2$  ppm can safely be used.

The variations in percentage mortality (Table 1) and LC<sub>W</sub>, LC j, and LC<sup>^</sup> values (Table 2) at different exposures indicated differential toxicity of DDVP to *Uia parsia* Verma *et at.* (1982) found the acute toxicity range of 6-10 ppm and 15-20 ppm

Table 2. Graphical analyfis : Acute toxicity value\* of DDVP\* to Liia parsia

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Exposure period (hr)	LCpt,	95* Fiducial limits		L Ci6	!! * *	Slope	95*
	(ppm)	Upper (ppm)	Lower (ppm)	Cppm)	(ppm)	function	confidence limit
24	1.015	1.324	0.778	0.468	2.175	2.157	1.305
48	0.750	1.005	0.560	0.295	1.950	2.571	1.340
72	0.554	0.698	0.440	0.282	1.072	1.950	1.260
96	0.482	0.606	0.384	0.272	0.851	1.768	1.256

•See composition in Table 1.

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for DDVP and malathion respectively on Saccobranchusfossilis. Their results proved that malathion is 2-3 times more toxic to fishes than DDVP. Pal (1983) reported 1.7 ppm and 12.9 ppm of DDVP as LC<sub>S</sub> and LC<sub>(S</sub> at 48 hr for *Tilapia mossambica*. According to Sailalha et al. (1981), the commercial grade malathion was 16.S times more toxic than the technical grade malathion on Tilapia mossambica. In the present bioassay study 0.75 ppm of commercial grade DDVP was found as 48 hr LC<sub>M</sub> for Liza parsia. Studies on acute toxicity of DDVP for brackishwater leleosts are not available hence data coufd not be compared with other results.

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