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Comparative regeneration of excised mantle tissue in one year and seven year old Indian pearl oyster, *Pinctada fucata* (Gould) grown under land-based culture system

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ABSTRACT

Excised mantle tissue (saibo) from the donor oyster is one of the important factors determining quality of cultured pearls. The present study was conducted to compare the process of regeneration of excised mantle tissue in one year and seven year old donor oysters, *Pinctada fucata* grown under land-based culture system. Menthol was used as relaxant prior to excision of mantle tissue, which was found to be effective at a concentration of 500 mg l⁻¹. The mantle tissue was found to regenerate within 3 months post-excision in both one year as well as seven year old *P. fucata*, with 100% survival. On gross examination, the regenerated mantle tissue of the 1 year as well as the 7 year old oysters appeared similar to that of the normal mantle tissue of the control group. Histological analysis demonstrated complete regeneration in 7 year old pearl oyster, *P. fucata*. The findings revealed that even aged donor oysters yielding good quality saibo can be recovered after mantle excision and could be further used as saibo donors for quality pearl production.

Keywords: Excised mantle tissue, Indian Pearl oyster, Mantle regeneration, Pinctada fucata, Saibo

Introduction

The major species of oysters used for cultured pearl production world over are the Akoya pearl oyster (Pinctada fucata, Gould), the silver/gold-lip pearl oyster (Pinctada maxima, Jameson) and the black-lip pearl oyster (Pinctada margaritifera, Linnaeus) from the family Pteriidae. Of these, P. fucata and P. margaritifera are of major importance with regard to availability and production of cultured round pearls as well as mabe pearls in India (Alagarswami, 1991; James et al., 1991; Dev and Durairaj, 1993; Kripa et al, 2007; Syda Rao, 2005; 2007). Studies were initiated on propagation of cultured round pearls about a century ago (Saville-Kent, 1893; George, 1966; Gervis and Sims, 1992). Cultured round pearls are produced by grafting a round nucleus and a piece of mantle tissue or 'saibo' from a sacrificed donor oyster into the gonad of a recipient oyster (Acosta-Salmon et al., 2004). Subsequent proliferation of the donor mantle tissue forms the pearl sac around the nucleus and continued deposition of nacre from the pearl sac onto the nucleus forms a cultured pearl (Gervis and Sims, 1992;

Acosta-Salmon *et al.*, 2004). The quality of cultured pearl depends greatly on the selection of appropriate donor oysters (Taylor, 2002)

Under normal circumstances, donor oysters are sacrificed during excision of mantle tissue. Recently, attempts have been made on in vivo regeneration of excised mantle tissue in different species of pearl oysters and the oysters were found to be able to survive after excision of mantle and were also found capable of complete regeneration of the excised mantle and its internal structures (Acosta-Salmon et al., 2004; Acosta-Salmon and Southgate, 2005, 2006; Mamangkey and Southgate, 2009). Even though, tissue/organ regeneration is a feature of every phyla in the animal kingdom, it varies among taxa and affected by several factors including age and cell complexity (Goss, 1969). So far, no attempt has been made to assess the in vivo regeneration of mantle tissue in pearl oysters grown for more than 5 years. The present work is a maiden attempt to evaluate the in vivo regeneration capability of the excised mantle tissue in response to age, in P. fucata grown under land-based culture system.

Materials and methods

The pearl oysters (P. fucata), used in this study were hatchery produced following the techniques of Alagarswami et al. (1983) and were cultured in land-based system (Syda Rao, 2001; 2007) at the Regional Centre of the Central Marine Fisheries Research Institute, Visakhapatnam, India. Briefly, the oysters were grown in specially designed concrete tanks having 10 t water holding capacity. They were maintained on the bottom of the tanks provided with suitable substratum to facilitate attachment of oysters. Pearl oysters were fed on a mixed diet of three species of algae viz., Chaetoceros calcitrans, Isochrysis galbana and Nanochloropsis salina by a specially designed drip method, so that the algal cell concentration in the tanks was continuously and automatically maintained at the desired rate (Syda Rao, 2001). They were gradually thinned as they grew, to a stocking rate of 60–70 nos. per m² by the time they reached implantation size. Growth recorded under land-based conditions was faster as compared to sea-based culture, owing to continuous availability of feed at required levels (Syda Rao, 2005). Mortality recorded under such land-based culture system was negligible.

Ten numbers each of 7 year old (7Y) oysters grown under land-based system as described above during the period 1998 to 2005, having a mean (± S.D.) dorso-ventral measurement (DVM) of 82.77 ± 12.68 mm and one year old (1Y) hatchery produced oysters with mean DVM of 56.17 ± 6.93 mm were used for the study. Studies in our laboratory have shown that menthol (-2-Isopropyl-5-methyl cyclohexanol, C₁₀H₂₀O) at a concentration of 500 mg l⁻¹ induced relaxations in P. fucata over a short period of time i.e., within 15 to 30 min of exposure allowing rapid recovery within 30 to 60 min post-excision, without any mortality. The same method was used in the present study prior to mantle excision. The oysters were anesthetized with 500 mg 1⁻¹ menthol for relaxation. Excision of mantle tissue was made from either the left or the right mantle lobe. The mantle tissue removed was approximately 5 mm x 20 mm in size. Following mantle excision, the two groups of oysters were returned to separate culture tanks immediately, maintained as described earlier and the survival was recorded over a period of 3 months post-excision. Control groups of 7Y oysters without mantle excision were also maintained under the same environmental conditions for comparison. The oysters were subjected to gross examination of the healed mantle, subsequently they were anaesthetized using menthol as described earlier and samples of mantle tissue were excised from normal 7Y oysters as well as from the regenerated mantle of both 7Y and 1Y oysters for histological examination. The fresh

tissue samples were fixed in Bouin's fluid for 24 h. They were then dehydrated in graded series of alcohol, embedded in paraffin, sectioned to 5 μ m thickness and stained with Periodic acid Schiff-Light Green (PAS-LG), Alcian blue-PAS and Azan as per the procedures described by Humason (1967) and Bancroft (1975).

Results

Application of menthol at a concentration of 500 mg l⁻¹ resulted in relaxation in 50% of oysters within 10 min and in the rest 30% within 15-30 min. There was relaxation without body collapse while mantle collapse was observed in 3% of the oysters, in the 7Y group. In both the treatment groups as well as in the control group, survival recorded was 100% during the period of 3 months observation. There was complete regeneration of the mantle within 58 days after mantle excision in 1Y oysters while it took 82 days to complete the process in 7Y oysters. Byssal secretion was noticed only in 1Y oysters while 7Y oysters had lost the power of byssal secretion even at the onset of the experiment. However, they were active in feeding.

Histological analysis of 7Y oysters showed complete regeneration of the mantle tissue (MT), 82 days post-excision of mantle. The regenerated and the normal MT showed typical bivalve mantle morphology with the marginal (with the inner, middle and the outer folds), pallial and central zones and secretion of conchiolin, in both 1Y and 7Y groups (Fig. 1, 2 and 3). The inner and outer surfaces of the mantle epithelium were well defined with ciliated epithelial cells. The inner fold of the regenerated mantle tissue was found to be larger in size than the other folds with large amount of muscle fibers. There were secretory cells in both inner and outer epithelia in the regenerated tissue (Fig. 3). The growth of tissue at the point of excision proceeded in the typical manner and there was no difference in the tissue aggregation and differentiation into respective layers of the regenerated MT of 1Y and 7Y groups. Both the longitudinal and radial muscles at the pallial zone were well developed in both the cases. There was no appreciable microscopic differentiation in the pallial zones, the outer epithelium of the secretory basophilic cells and the mucous secretory cells, either between the regenerated mantle of 1Y and 7Y groups or between the normal MT and regenerated MT of 7Y oysters. There was no abnormality observed in the anatomy of the mantle regeneration process in 7Y oysters as compared to 1Y oysters. However, the prismatic layer appeared darker in 1Y MT than that of the 7Y MT. The new nacre secreted on the inner surface of the shell adjacent to the wound area appeared thinner and irregular in 7Y regenerated oysters as compared to that of 1Y group and control 7Y group.

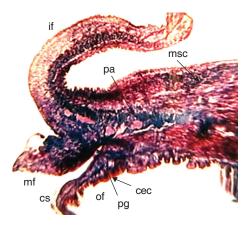


Fig. 1. Histological section showing regenerated mantle folds in 1 year old *Pinctada fucata*, 58 days after mantle excision (Azan; X 200).

pa: artery, cec: columnar epithelial cells, msc: muscle, cs: conchiolin secretion, if: inner fold, mf: middle fold, of: outer fold, pg: periostracal groove.

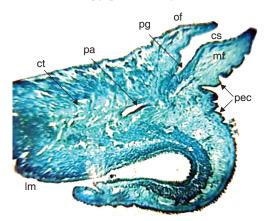


Fig. 2. Photomicrograph of normal mantle tissue of 7 year old *P. fucata* (PAS-LG; X 200)

if: inner fold, mf: middle fold, of: outer fold, cs: conchiolin secretion, pec: pigmented epithelial cells, lm: longitudinal muscles, ct: connective tissue, pa: pallial artery, pg: periostracal groove.

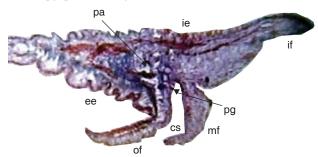


Fig. 3. Histological section of regenerated mantle tissue of 7 year old *P. fucata*, 82 days after mantle excision (Alcian blue-PAS; X 100).

if: inner fold, mf: middle fold, of: outer fold, cs: conchiolin secretion, ee: external epithelium, ie: internal epithelium, pa: pallial artery, pg: periostracal groove

Discussion

Relaxants have been used with pearl oysters to facilitate internal inspections, seeding operations for pearl production and also to obtain saibo tissue from the donor oysters (Norton et al., 2000; O'Connor and Lawler, 2003; Acosta-Salmon et al., 2004; Acosta-Salmon and Southgate, 2005). The use of an appropriate relaxant facilitates removal of saibo tissue from live oysters without killing the oysters. A number of chemicals have been used to relax pearl oysters with varying success and limitations (Norton et al., 1996; Acosta-Salmon and Rangel-Devalos, 1997; Saucedo et al., 2001; O'Connor and Lawler, 2003). Norton et al. (2000) evaluated the quality of pearls produced using relaxed oysters. However, they did not use relaxed mantle tissue as saibo. Acosta-Salmon et al. (2004) used 500 mg 1-1 benzocaine for relaxing *P. fucata* with appreciable results. During the present study, it was found that menthol at a concentration of 500 mg l⁻¹ can be used safely for relaxation of both 7Y as well as 1Y oysters. There was relaxation without body collapse while mantle collapse was observed in 3% of the oysters, mostly in the 7Y group, which is quite acceptable when the number of oysters taken into account is high. Acosta-Salmon et al. (2005) opined that oysters showing body collapse but no mantle collapse could still be used as saibo donors. Tranter (1957) used menthol crystals on P. maxima and Dev (1994) used powdered menthol in P. fucata. But so far, effective concentration of menthol has been scarcely reported for relaxation of oysters without mantle collapse.

Our studies demonstrated the regeneration process of excised mantle tissue in 7Y and 1Y oysters and also the morphological similarity between regenerated mantle tissue and the normal mantle tissue in 7Y oysters. The regeneration process was found to be complete within 3 months as indicated by the restoration of physical as well as functional tissue structure characterized by the three mantle folds, secretary cells, muscle fibres and conchiolin secretion. Similar results were reported in young *P. fucata* by Acosta–Salmon and Southgate (2005). However, Carlson (2007) reported incomplete restoration of mantle tissue possibly caused by nutritional or other types of deficiency.

Gervis and Sims (1992) reported that the age of a pearl oyster influences the speed of mantle regeneration. For cultured pearl production, older pearl oysters are generally preferred to be used as saibo donors than young ones owing to their slower growth rate. The saibo tissue from such oysters secrete nacre at a slower rate resulting in better quality pearls (Grevis and Sims, 1992). Garcia-Gasca *et al.* (1994) reported that large secretory basophilic cells play an important role in nacre layering. From the fact that in 7Y oysters the mantle tissue took about 24 days more to grow back to its normal size than that of 1Y oysters, it is quite evident that mantle regeneration is rather a normal process but the time of recovery may differ in accordance to the age of the oyster. This may be attributed to the difference in efficiency in the nutrient storage and utilization. Studies have shown that mantle tissue functions as a site for storage of nutrients in bivalves (Barber and Blake, 1981; Mathieu and Lubet, 1993; Pekkarinen, 1994; Acosta-Salmon et al., 2004). Carlson (2007) stated that nutritional deficiency influences the mantle regeneration process. In the land-based culture conditions as used in the present investigation, the physico-chemical environment and the nutrient availability were kept nearly optimal (Syda Rao, 2007). This might have lead to early mantle regeneration in 1 Y oysters *i. e.*, in 58 days as compared to that of 60 days or more (Acosta-Salmon and Southgate, 2005) recorded under sea based culture conditions. Further, the size of excised mantle tissue also influences the time required for complete healing.

Following mantle excision, pearl oysters are susceptible to the ambient environmental conditions. It has been observed that the feeding process slows down up to 2 days after mantle excision. This may be due to significant loss of mantle tissue or due to the excision shock. Earlier studies have recognized that the rate of wound healing in pearl oysters is likely to be influenced by their physiological state (Acosta-Salmon and Southgate, 2005; 2006) and by seasonal patterns of nutrient storage which have been described for marine bivalves including pearl oysters (Saucedo et al., 2002). Mamangkey and Southgate (2009) found that wound healing with epithelialization was completed within 3-4 days of mantle excision while morphogenesis took 3 months. Carlson (2007) stated that complete regeneration requires all cellular functions to be in place and this process takes much longer than the wound healing. Acosta-Salmon and Southgate (2005) reported that following the mantle excision, both the ends rolled inwards to the centre reducing the size of wound, minimizing hemorrhaging. Regeneration was rapid and complete, once sealing of wound has occurred. Similar pattern was reported during siphon regeneration in Scrobicularia plana (Hodgson, 1982) and in Macoma balthica (Pekkarinen, 1994). Acosta-Salmon et al. (2004) opined that damage of mantle tissue and the energetic cost of the healing and regeneration process do not interfere with other body functions. They reported the regeneration of mantle tissue in vivo in P. fucata cultured in long lines and have shown that oysters used as saibo donors need not be killed and could be used as future brood stock and as future saibo donors. Maintaining a pool of 3-4 year old population will be promising for continuous availability of desired saibo of suitable quality.

The present study is the first description of *in vivo* mantle regeneration in 7 year old pearl oyster, *P. fucata*. The results indicated that oysters can regenerate the excised

mantle tissue in less than three months irrespective of age under land-based culture conditions. Though, a few of the earlier studies have reported regeneration of mantle tissue in pearl oysters, until now no study has been done on comparative regeneration process of mantle tissue with respect to age. The findings of the present study make a significant contribution to our knowledge on tissue regeneration process in pearl oysters with respect to age. As the breeding efficiency declines sharply after 4 years of growth in P. fucata (Syda Rao, 2007), oysters above 4 years producing high quality pearls could possibly be used as saibo donors repeatedly, without sacrificing the animal. Demonstration of regeneration of MT even in the aged donor in vivo under captive conditions in the present study indicate that a pool of high quality donor oysters can be maintained as future saibo donors. The regeneration capacity of the excised mantle tissue, also makes it possible to make use of good quality saibo donors in selective breeding programmes.

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