

Recurrent dinoflagellate blooms in the South-eastern Arabian Sea -A preliminary assessment with focus on *Ornithocercus magnificus* blooms along the nearshore waters off Dakshina Kannada, South-west coast of India

LAVANYA RATHEESH^{1,2}, PRATHIBHA ROHIT³, BINDU SULOCHANAN³ AND P. PRANAV¹

¹ICAR-Central Marine Fisheries Research Institute, P.B. No. 1603, Kochi - 682 018, Kerala, India

²Mangalore University, Mangalagangotri, Mangalore - 574 199, Karnataka, India

³Mangalore Research Centre of ICAR-Central Marine Fisheries Research Institute, Mangalore - 575 001, Karnataka, India e-mail: lavanyacmfri2010@gmail.com

ABSTRACT

Frequent bloom incidences of the dinoflagellate *Ornithocercus magnificus* Stein, 1883 along the surface waters off Southeastern Arabian Seas (SEAS) have been reported since 2015 and specifically during the years 2015, 2019 and 2020. All these blooming incidences coincided with the strong El Nino phases in the Indian Ocean. The present study addressed the bloom $(1.65 \times 10^6 \text{ cells } 1^{-1})$ of this species in the nearshore waters (7 m depth) off Dakshina Kannada (Surathkal and Chitrapur) on 11.02.2016 which occurred during pre-monsoon period and corresponding to the super El Nino year 2015/2016. A detailed evaluation of the physico-chemical characteristics and phytoplankton abundance as well as community structure was made, concurrent to the three phases *viz.*, pre-bloom, bloom and post-bloom periods in this region. The study revealed that the bloom occurred in high saline (35.0 ± 0.91 PSU), well oxygenated (7.414 ± 0.823 mg 1^{-1}) and nitrogen limited waters (oligotrophic conditions) and had a positive correlation with oceanic nino index (ONI; $r_s = 0.790$, p<0.001). Some of the *O. magnificus* (3%) harboured ectosymbionts probably cyanobacteria in their cingulum in response to the beginning of a stratified oligotrophic condition in this region. The water quality was fair during the bloom period with no conspicuous discolouration of the surface waters. The present study also attempted to evaluate the influence of the increasing frequency of the *O. magnificus* blooms on the oilsardine fishery along the SEAS.

Keywords: Cyanobacteria, Dinoflagellate, Ectosymbionts, Nitrogen limited waters, Oilsardine, Pre-monsoon season, SEAS, Sea surface temperature (SST)

Introduction

Ornithocercus sp., belonging to the family Dinophysaceae and characterised by complex structure is an attractive planktonic dinoflagellate represented by 15 species globally (Cleve, 1900, 1901, 1903; Kofoid, 1907; Kofoid and Skogsberg, 1928; Wood, 1963, 1968; Gomez, 2005). In the Indian Ocean also their occurrence has been reported by Taylor (1976) and Wood (1963) identifying 9 and 8 species of this genus respectively. Kuzmenko (1975) has recorded the presence of three species belonging to this genus from the northern Arabian Sea. Furthermore, Syed *et al.* (2008) identified 5 species of this genus from the surface tropical waters of northern Arabian Sea shelf off Pakistan, of which four were new records.

Earlier Lewis (1965) reported a considerably high abundance of *Ornithocercus magnificus* Stein, 1883 (1.175 x 10^5 cells l⁻¹⁾ in association with a red tomato soup bloom of *Gonyaulax polygramma*,off Arabian Sea at 10 m depth during the post-monsoon season. High densities of the species in the Arabian Sea, has also been addressed by Subrahmanyan (1958) indicating discolouration of surface waters, though the term "bloom" was not mentioned.

Other than these reports, blooms of this genus in the surface waters of Arabian Sea have been a rare occurrence. However, in the recent decades, blooms of this genus have been reported frequently from the northern Indian Ocean. Karthik *et al.* (2017) reported the bloom of *O. magnificus* in the coastal waters (surface) off Karnataka during the pre-monsoon of 2015. In addition, Lavanya *et al.* (2020), have also reported on the blooms of *O. magnificus* in the inshore surface waters, off Kochi, Kerala during the post-monsoon season of 2019. Interestingly, after the subsidence of the bloom in 2019 and within a gap of one month, the species was observed once again in high abundance off Kochi inshore waters during February 2020, with presence of ectosymbionts harboured in the crown of the cells.

O. magnificus is used as an indicator species for climate and environmental change along the South Korean waters (Kim *et al.*, 2008). Korea being in the temperate region, the warmest water temperature recorded

was 23.7°C, however, the presence of 19 new tropical dinoflagellate species, including the indicator species O. magnificus in very high abundance, indicated a significant change in the dinoflagellate composition in Korean waters. These observed changes were attributed to warming up of the coastal waters (Kim et al., 2008). Kim (2013) reported the presence of 34 as yet unrecorded dinoflagellate species from South Korean waters and 23 of these were tropical species. Though the presence of O. magnificus in the Indian tropical waters is common and generally not considered as harmful, its high abundance in the surface waters has been linked to increasing sea surface temperatures (SST) (Lavanya et al., 2020) and consequent environmental changes. Unlike blooms of Trichodesmium sp., which are common during the premonsoon season, frequent blooms of Ornithocercus sp. along the surface waters of Arabian Sea along the SEAS is rare. In a span of 5 years (2015-019), the frequency of occurrence of O. magnificus blooms along the west coast of India has been increasing (Karthik et al., 2017; Lavanya et al., 2020) indicating a discernible change in the marine ecosystem especially in terms of increased SST which in turn could have negative impacts on primary productivity and the marine food web (John et al., 2018). Furthermore, the presence of this species in the gut of oilsardine (caught by bottom gillnets and bottom trawlnets during the pre-monsoon), also leads to a point of concern on the possible cause of the blooming of this bottom dwelling species in the surface waters (Kagwade, 1964; Dulkhed et al., 1972). Thus, mere occurrence of the bloom or even a higher abundance of this species needs to be extensively evaluated. Even though Karthik et al. (2017), did not report any correlation of the bloom of O. magnificus with SST, the bloom of this species occurred during pre-monsoon concurrent to the El Nino year (2014-15). The present bloom also occurred during the pre-monsoon of a very strong super El Nino year (2015-16).

The present study investigated the variability in the physico-chemical attributes of the nearshore surface waters, off Dakshina Kannada, south-west coast of India, during three phases of *O. magnificus* bloom that occurred on 11.02.2016 and probed in to the increasing frequencies of *O. magnificus* blooms along the SEAS.

Materials and methods

Study area

Two nearshore stations, Surathkal (13°0'23.882"N; 74°46'43.097"E; 6-7 m depth) and Chitrapur (12°56'32.761"N; 74°47'25.56"E; 7 m depth) off Mangaluru in Dakshina Kannada District were monitored (Fig. 1). Seawater and phytoplankton samples were collected on a monthly basis.

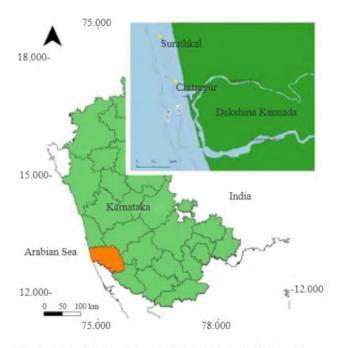


Fig. 1. Map showing the sampling locations in the nearshore waters off Dakshina Kannada on 11.02.2016

Sample collection and data analysis

Collection and analysis of water samples

Surface seawater samples were collected during the early morning in polypropylene bottles and brought to the laboratory for further analysis of their physicochemical characteristics. Surface water temperature was measured using a centigrade mercury thermometer (-10°C-110°C; OMSONS). Salinity was measured potentiometrically using a multi-parameter instrument (WTW 320i, Xylem Analytics LLC, Germany). For dissolved oxygen estimation, water samples collected in 125 ml BOD bottles were immediately fixed with Winkler reagents and subsequently analysed following Winkler's method (Strickland and Parsons, 1968). Chlorophyll a was estimated by filtering the water samples using GF/F Whatman glass fibre filters of pore size 0.7 µm followed by pigment extraction using 90% acetone and estimation in spectrophotometer (UV-VIS Lambda 365, Perkin Elmer). Dissolved inorganic nutrients (ammonia, nitrite, nitrate, phosphate and silicate) were determined using standard analytical methods (APHA, 1981). Water quality indices (WQI) were also evaluated, as per the selected environmental indicators of USEPA (2012).

Phytoplankton sample collection and analysis

Phytoplankton samples were collected by horizontal hauling of a standard plankton net of mesh size 20 μ m and 0.5 m² mouth diameter, at a speed of 2 knots for 10 min and preserved using 4% formaldehyde. Enumeration and

identification of the phytoplankton samples were carried out following the standard phytoplankton identification manuals (Subrahmanyan, 1958; 1968) under light microscope (LEICA ICC50).

Statistical analysis

The Shannon and Weiner index was estimated to determine the phytoplankton species richness and diversity in the bloom (Shannon and Weaver, 1949) using the statistical software PRIMER (Ver.6.3.2). Spearman correlation analysis (2-tailed) was also carried out using SPSS software (SPSS Statistics 23) to evaluate the significant relation of various abiotic variables with the abundance of *O. magnificus*.

Secondary data

To understand the intensity of El Nino and La Nina events, oceanic nino index (ONI) data pertaining to 2015-2019 years were accessed from the National Oceanic and Atmospheric Administration (NOAA). Monthly average rainfall data during the study period was collected from Indian meteorological department. Monthly fish catch data for the period was taken from data bank of National Marine Fisheries Data Centre (NMFDC) of ICAR-Central Marine Fisheries Research Institute (ICAR-CMFRI).

Results and discussion

Bloom characteristics

Microscopic examination of the phytoplankton sample collected on 11.02.2016 from the nearshore waters off Surathkal (1.65x106 cells 1-1) and Chitrapur (7.6x105 cells 1-1) in Dakhsina Kannada revealed O. magnificus as the dominant phytoplankton species responsible for the bloom (Fig. 2). The incidence of this genus in the Arabian Sea with exclusive occurrence restricted to pre-monsoon phase has been reported by Subrahmanya (1958), Kuzmenko (1975), Gomez (2005), Syed et al. (2008) and Karthik et al. (2017), with an exception on their occurrence during post-monsoon by Lavanya (2020). The length of O. magnificus ranged from 70-100 µm with 6-8 complete ribs (Wood, 1954, 1968; Taylor, 1976). Coinciding with the earlier reports, the present bloom also occurred during the pre-monsoon period. In order to have a detailed seasonal evaluation of the bloom, the occurrence and abundance was monitored and categorised as Pre-bloom, Bloom and Post-bloom phases. The pre-bloom sampling was conducted on 25.01.2016 and were characterised by a cell count of 6.6 x103 cells 1-1. A spike in their abundance was noticed in February reaching to a maximum abundance (1.65 x 10⁶ cells 1⁻¹) and hence was categorised as bloom period. Following the bloom period, a subsequent decline in their abundance reaching to zero was noticed during March 2016 and hence was categorised as the post-bloom phase. There was no conspicuous discolouration of the surface waters during the sampling periods, even during the bloom, though Subrahmayan (1958) had mentioned about the discolouration of waters associated with high densities of this species. Sighting of marine birds such as seagulls and terns during the bloom period indicated evidence of fish availability in the region.

Water quality and physico-chemical characteristics during the bloom

The physico-chemical characteristics differed during the three distinct phases of the bloom (Fig. 3). The bloom period was notable for significantly higher salinity (35.2±0.91 PSU) (p<0.05) and oxygenated waters (p<0.05) with dissolved oxygen concentration of 7.414±0.823 mg 1-1, higher than the pre-bloom and post-bloom phases. The SST during the bloom ranged from 30 to 30.5°C and was relatively higher than the pre-bloom and lower than the post-bloom phase. The average chlorophyll a concentration was observed to be 3.382±0.53 mg m⁻³ ranging between 2.72 to 3.8±0.53 mg m⁻³ with lowest observed during the pre-bloom phase. Inorganic phosphate was in the range 0.03-0.04±0.01 mg l-1, while nitrate $(0.003\pm0.01 \text{ mg } l^{-1})$ and nitrite $(0.005\pm0.01$ mg 1-1) concentrations were observed to be low during the bloom revealing that the waters were No- limited, indicating the beginning of an oligotrophic condition. Tarangkoon et al. (2010) recorded occurrence of symbiont-bearing dinoflagellates in oligotrophic water masses with low nutrient (N-limited) and chla concentration. The average ammonia concentration was observed to be 0.065±0.045 mg l⁻¹, while the ammonia concentration showed a sharp decline from the pre-bloom phase to almost nil during the post-bloom phase. The maximum ammonia concentration (0.11±0.045 mg l-1) observed during the pre-bloom phase was not above the permissible



Fig. 2. *O. magnificus* collected during the bloom observed on 11.02.2016 in the nearshore waters of Dakshina Kannada

Occurrence of Ornithocercus magnificus blooms along the south-west coast of India

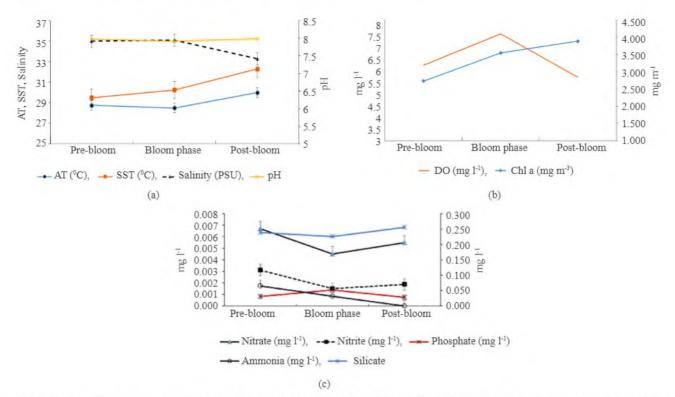


Fig. 3. Physico-chemical parameters during the different phases of the bloom of *O magnificus* off Dakshina Kannada. (a) Air temperature (AT), (b) Seasurface temperature (SST) and (c) Salinity (PSU)

limit for coastal waters, hence fish mortality was also not observed. This also indicates that other phytoplankton species which could not survive or tolerate lower nutrient levels or an oligotrophic condition in the ambient waters started degrading. There was a marked decrease in the diatom abundance during the post-bloom phase. The water quality index as per USEPA (2021) rating was fair during the bloom. The variation in physico-chemical parameters was also studied for the years from 2014 to 2016 during the months from January to March (Fig. 4).

Phytoplankton community structure

O. magnificus dominated with a contribution of 54-99% of the total phytoplankton population at all stations, during the pre-bloom and bloom phases and declined gradually to a complete absence during the post-bloom. The abundance of *O. magnificus* during the pre-bloom, bloom and post-bloom phases is represented in Fig. 5.

Analysis of phytoplankton samples revealed the presence of 53 species, majority of them being diatoms (32 species), followed by dinoflagellates (21 species) and blue-green algae (1 species) (Fig. 6). A similar trend of species richness (S) was observed during the pre-bloom as well as peak bloom period. Of the 32 diatom species identified, 11 species were recorded during the pre-bloom,

9 species during the bloom phases, while 5 species were recorded during the post-bloom period. Similarly, of the 21 dinoflagellates species recorded, 14 species were observed during the pre-bloom and bloom periods and 11 species during the post-bloom period. The Shannon Weiner diversity index (H') was lowest during the bloom phase [H'(log2) = 3.37], while the pre-bloom phase recorded highest value [H'(log 2) = 2.55]. During the pre-bloom phase, O. magnificus rapidly started proliferating and reached peak abundance leading to the bloom and almost displaced the diatom population as this dinoflagellates are adapted to survive and thrive well in low nutrient conditions. This was followed by a sudden decline in the abundance of O. magnificus (post-bloom phase) due to the sudden drop in salinity from 35 to 32 PSU respectively as a result of the unexpected rainfall (7.53 mm).

During the pre-bloom and bloom periods, Coscinodiscus spp. dominated among diatoms, followed by Hemiaulus hauckii, Surirella spp., Biddulphia mobiliensis and Navicula spp. Among the dinoflagellate community, Ornithocercus magnificus was dominant followed by Dinophysis miles, Protoperidinium spp., Ceratium furca, Ceratium tripos, Dinophysis caudata and Pyrophacus spp. The post-bloom phase was notable for the presence of Trichodesmium spp. indicating a state of extended stratified condition. Lavanya Ratheesh et al.

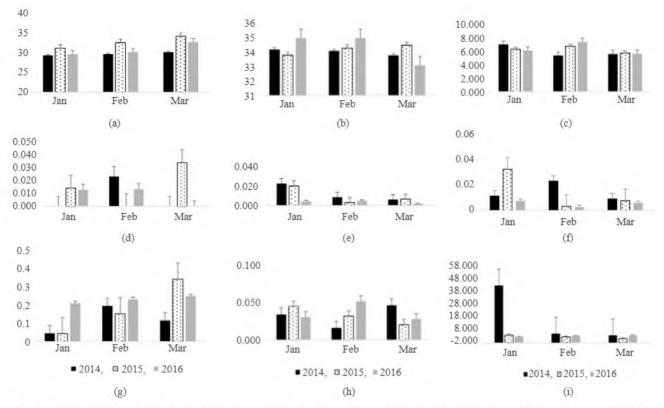


Fig. 4. Variations in the physio-chemical parameters of the nearshore waters off Dakshina Kannada during January-March of 2014-2016.
(a) Seasurface temperature (SST (⁰C), (b) Salinity (PSU), (c) Dissolved oxygen (DO mg l⁻¹), (d) Ammonia (mg l⁻¹), (e) Nitrite (mg l⁻¹), (f) Nitrate (mg l⁻¹), (g) Silicate (mg l⁻¹), (h) Phosphate (mg l⁻¹) and (i) Chlorophlla (Chl a, mg m⁻³)

Ectosymbiont bearing O. magnificus

Microscopic examination (40x) of the preserved individuals of *O. magnificus* (Fig. 7a,b) revealed the presence of ectosymbionts, possibly cyanobacteria in the cingulum as phaesosomes of some cells (3%) along with some remnants that resembled the ectosymbiont, in their food vacuoles. Similar observation of ectosymbionts was reported by Jyothibabu *et al.* (2006); Farnelid *et al.* (2010) and Tarangkoon *et al.* (2010)., in live specimens of *O. magnificus.* (Fig. 7a,b). Further detailed examination by transmission electron microscopy (TEM) as done by

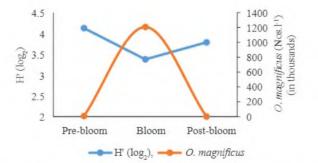


Fig. 5. *O. magnificus* abundance along the nearshore waters off Dakshina Kannada during the bloom

Lucas (1991); Farnelid *et al.* (2010) and Tarangkoon *et al.* (2010) could not be carried out during the present investigation.

Heterotrophic dinoflagellates like *O. magnificus* that lack photosynthetic pigments, are known to exclusively feed through osmotrophy (Droop, 1974). However, they have been observed to provide favourable microenvironments for free floating cyanobacteria, for efficient nitrogen fixation when the upper euphotic waters are stratified and the water masses become nitrogen limited (Gordon *et al.*, 1994; Jyothibabu *et al.*,

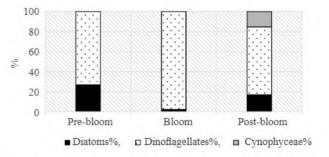


Fig. 6. Phytoplankton composition during the three phases of the bloom along the nearshore waters off Dakhsina Kannada

Occurrence of Ornithocercus magnificus blooms along the south-west coast of India

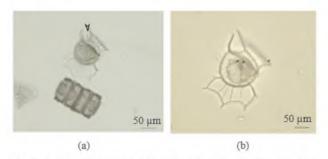


Fig. 7. Light microscopy of preserved cells of *O. magnificus* showing (a) Ectosymbionts in the cingulum (arrow head) and (b) Enlarged view showing the presence of ectosymbionts in the food vacuoles (arrows)

2006; Farnelid et al., 2010 and Tarangkoon et al., 2010). Although Trichodesmium sp. plays a major role in the nitrogen cycling of the Arabian Sea, their abundance got higher only during the extended stratified conditions in post-bloom phase. During the initial stages of thermal stratification, the primary productivity is contributed by the free floating minute cyanobacteria (Li et al., 1995; Maranon et al., 2003; Jyothibabu et al., 2006). The stable chlorophyll a and dissolved oxygen concentrations (r = 0.985, p < 0.001) observed could be attributed to the presence of the ectosymbionts, resembling cyanobacteria enabling the host dinoflagellate to survive in oligotrophic waters through mixotrophy. The phaesosomes also have an advantage harbouring these dinoflagellates as they get protected from high oxygenated waters that can inactivate their N2 fixing nitrogenase enzyme activity (Paerl et al., 1987; Jyothibabu et al., 2006).

O. magnificus surface blooms and its possible link to high SST

Phytoplankton are ideal indicators of any sudden environmental perturbations as they have a high turnover rate and immediate biological response (Racault et al., 2017). The SST along the west coast of India, normally shows a double oscillation. The first maximum is during the pre-monsoon months (April-May) and the second maximum during the beginning of post-monsoon season (Subrahmanyan, 1959). Similar trend was also observed in the salinity distributions along this coast. Under normal conditions, by the end of the pre-monsoon season, the dry winds push the warmer surface waters from the coast and bring in the deeper, cold nutrient rich waters upward, leading to coastal upwelling. But during strong El Niño events, these strong winds weakens and fails to displace the existing warm waters which gets transported as Kelvin waves leading to a delay in the coastal upwelling, resulting in thermal stratification and oligotrophic conditions (John et al., 2018)

The present report on the bloom of *O. magnificus* that occurred during El Nino years is similar to earlier reports

on its incidence along the SEAS. Impacts of El Nino, especially on primary productivity and phytoplankton populations have been found to be greatest in the tropical and subtropical regions of the world. Furthermore, the 2015/2016 El Nino as in the case of the present bloom incidence, happened to be one of the strongest and super El Nino event (Weare 1979; Saji et al., 1999; Bo Lu et al., 2018) in the 20th century breaking the warming record in the central Pacific, reaching its peak of 3°C in November 2015, represented by the NINO3.4 and NINO4 indices. after the 1982/83 and 1997/98 events. According to Saji et al. (1999) and Bo et al. (2017), the Indian Ocean SST during pre-monsoon months of 2016, was passively modulated by this 2015/2016 super El Nino leading to excess warming up of the Indian Ocean. This modulation led to positive SST anomalies in both western and eastern IOD (Indian Ocean Dipole) during January to April 2016 (Fig. 8). Kripa et al. (2018), also have reported a noticeable 1.1°C increase in the average SST from 28.6°C, in the sardine habitats off Kochi waters during 2015/2016 El Nino event. The tropical Indian Ocean also was dominated by positive SST anomalies exceeding 0.6°C during this period. The ONI index was also highest during pre-bloom and bloom periods (2.1). In the present study, the SST of pre-monsoon season of 2015 and 2016 was also observed to be higher as compared to 2014 (Fig. 4).

Warm waters with higher salinity are reported to be conducive for the growth of dinoflagellates (David et al., 1960; Dae et al., 2004). In the present study, O. magnificus abundance showed highly significant positive correlation with ONI index (r = 0.790, p<0.001), significant correlation with salinity (r = 0.689, p<0.05) and dissolved oxygen values (r = 0.713, p<0.05) while it was negatively correlated with nitrate concentration $(r_{s} = -0.646, p < 0.05)$ (Table 1). ONI index was also found to be negatively correlated with chlorophyll a (r = -0.688, p<0.05). Though O. magnificus abundance did not show any direct correlation with SST, its significant correlation with ONI index attributes its tolerance to increasing SST. Ectosymbiont bearing O. magnificus was found to be abundant at temperature range of 16.5-30.1°C in the upper 100 m water column of the Indian Ocean (Tarnagkoon et al., 2010). The SST (30±1.31°C) during the bloom period was higher than the range reported earlier. Syed et al. (2008) had observed maximum (16.47%) frequency of O. magnificus along the North Arabian Sea when the water temperature ranged between 23.04 and 28.2°C. Surface salinity (35.0±0.91 PSU) was comparatively lower than that reported by Syed et al. (2008). The salinity during the pre-monsoon season ranged from 36.3 to 36.5 PSU. Such environmental conditions helped the ectosymbiont bearing dinoflagellates to exhibit mixotrophy which can be defined as the combined use of photosynthetic and

60E 80E 100E 60E 80E 100E 60E 80E 100E 60E 80E 100E °C -2 -1.5 -1 -0.8 -0.6 -0.4 -0.2 0 0.2 0.4 0.6 0.8 1 1.5 2

Fig. 8. SST anomalies in the Indian Ocean over the period from 1989-2021 (http://www.cpc.ncep.noaa.gov/products/GODAS/)

			Air Temp	SST	pН	Salinity	Silicate	Phosphate	Nitrate	Ammonia	Dissolved Oxygen	Chla	ONI Index	O, magnificus
	Air Temp	Correlation Coefficient	1.000	.692*	-087	.439	.549	102	075	370	146	.307	.150	.222
		Sig. (2-tailed)		0.27	.812	.204	.100	.778	.836	.293	.687	.387	.679	.537
		N	10	10	10	10	10	10	10	10	10	10	10	10
	SST	Correlation Coefficient	.692*	1.000	.242	013	.383	028	229	385	034	.494	252	.080
		Sig. (2-tailed)	.0.27		.501	.973	.275	.939	.525	.272	.925	.147	.482	.826
		N	10	10	10	10	10	10	10	10	10	10	10	10
	pH	Correlation Coefficient	087	.242	1.00	790**	243	813**	.139	072	241	.379	711*	559
		Sig. (2-tailed)	.812	.501		.007	.498	.004	.702	.960	.485	.280	.021	.093
		N	10	10	10	10	10	10	10	10	10	10	10	10
	Salinity	Correlation Coefficient	.439	013	790**	1.000	.431	.471	184	.204	272	- 229	.803**	.689*
		Sig. (2-tailed)	.204	.973	.007		.214	.170	.610	.571	.446	.524	.005	0.27
		N	10	10	10	10	10	10	10	10	10	10	10	10
	Phosphate	Correlation Coefficient	- 102	028	813**	.471	.183	1.000	096	.006	227	- 264	.404	.360
		Sig. (2-tailed)	.778	.939	.004	.170	.613		.792	.987	.528	.461	.247	.307
		N	10	10	10	10	10	10	10	10	10	10	10	10
	Nitrate	Correlation Coefficient	- 275	229	.139	- 184	080	096	1_000	152	889**	.505	561	646
		Sig. (2-tailed)	.836	.525	.702	.610	.826	.792		.676	.001	.137	.092	0.44
		N	10	10	10	10	10	10	10	10	10	10	10	10
	Ammonia	Correlation Coefficient	370	385	018	.204	.293	.006	152	1.000	.368	405	.328	.104
		Sig. (2-tailed)	.293	.272	.960	.571	.412	.987	.676		295	.246	.355	.776
		N	10	10	10	10	10	10	10	10	10	10	10	10
	Dissolved	Correlation Coefficient	146	034	251	.272	098	.227	889**	.368	1.000	681	.694"	.713*
	Oxygen	Sig. (2-tailed)	.687	.925	.485	.446	.789	.528	.001	.295		.0.30	0.28	0.21
		N	10	10	10	10	10	10	10	10	10	10	10	10
	Chla	Correlation Coefficient	.307	.494	.379	229	.024	264	.505	405	681*	1.000	688*	250
		Sig. (2-tailed)	.387	.147	.280	.524	.947	.461	.137	.246	.030		0.28	.486
		N	10	10	10	10	10	10	10	10	10	10	10	10
	ONI Index	Correlation Coefficient	.150	252	711*	.803**	.169	.404	561	.328	.694*	668*	1.000	.790**
		Sig. (2-tailed)	.679	.482	.021	.005	.640	.247	.092	.355	.026	.028		.006
		N	10	10	10	10	10	10	10	10	10	10	10	10
	0.	Correlation Coefficient	.222	.080	559	.689*	018	.360	646*	.104	.713*	- 250	.790**	1.000
	magnificus	Sig. (2-tailed)	.537	.826	.093	.027	.960	.307	.044	.776	.021	.486	.006	
		N	10	10	10	10	10	10	10	10	10	10	10	10

*Correlation is significant at the 0.05 level (2-tailed) **Correlation is significant at the 0.01 level (2-tailed)

GODAS SST Anomaly (5 m) (8°N - 12°N), Indian Ocn

heterotrophic nutrition within a single organism (Kofoid and Skogsberg, 1928; Lucas, 1991; Gordon *et al.*, 1994, Jyothibabu *et al.*, 2006). This mode of nutrition helps these dinoflagellates to thrive in the oligotrophic conditions and to dominate over other phytoplankton communities. High surface salinity caused due to evaporation during the pre-monsoon phase coupled with warm oligotrophic waters, with the help of the ectosymbionts could have favoured the formation of *O. magnificus* bloom.

Significance of Ornithocercus sp. bloom in fisheries

Kagwade (1964), reported that among the 6 dinoflagellate species found in the gut of oilsardine, the monthly frequency of occurrence of Ornithocercus sp. ranged between, 6.6 and 28.5% during pre-monsoon months and post-monsoon months when SST and salinity in the Arabian Sea was maximum. Oilsardines are surface feeders and are usually exploited by purse-seines, cast net, boat-seines and surface gillnets operating at depths ranging from 1 to 5 m and by drift gillnets and pelagic trawlnets operating nearshore between 10-20 m depths. However, Dulkhed (1972) observed difference in the food composition of oilsardine caught by gillnets and trawl nets operating along Dakshina Kannada during the pre-monsoon season. Oilsardine from the surface waters showed poor feeding and the diet mainly consisted of Coscinodiscus spp. while, sardines caught from the bottom waters had gorged stomachs and the diet mainly (97%) comprised of bottom dwelling Ornithocercus sp. Coincidently, this report based on the study made during 1963-64 also happened to be an El Nino year. Tasaduq et al. (2019) also observed the presence of Ornithocercus sp. as one of the main food of oilsardine caught using trawlnets during the pre-monsoon season during 2010-12 along the Ratnagiri coast, off Maharashtra. These reports clearly indicated that the Indian oilsardine occasionally foraged heavily on Ornithocercus sp., by moving to the bottom waters during the pre-monsoon seasons. When Kelvin waves lower the thermocline and brings a situation where the bottom waters becomes warm, a favourable environment is created for the temperature tolerant species like Ornithocercus sp. which proliferate with the help of its ectosymbionts and finally outnumber other phytoplankton species. The visualisation of depth-wise temperature anomalies of 2015-2016 El Nino event from January 2015 to December 2016, published by NASA (National Aeronautics and Space Administration) clearly depicts the temperature anomalies during the bloom period (Fig. 9). This visualisation also confirmed the warming of the bottom waters during the beginning of the bloom event. These events in turn impacted the feeding habits of oilsardine which usually migrate to the bottom waters for feeding during this season.

A decline in oilsardine catch during El Nino events has been reported by Rohit et al. (2018) and Shetye et al. (2019). They observed a collapse in oilsardine fishery from 1.55 lakh t in 2014 to 0.46 lakh t during the strong 2015-16 El Nino event along the Kerala coast. The catch recovered (1.27 lakh t) in the following year (2017), which was a normal ENSO (El Nino-Southern Oscillation) year. The 2015-16 El Nino event had a negative impact on the oilsardine fishery, which in turn resulted in high SST and reduced upwelling velocity, leading to minimum cooling of the surface waters and thereby favouring the dominance of nano and pico phytoplankton. A similar situation was observed along the coast of Karnataka, where the oilsardine fishery dropped down from 1.43 lakh t (2014) to 0.29 lakh t in 2015 and 0.27 lakh t in 2016 (Fig. 10.) In addition to the decline in catch Rohit et al. (2018) also reported on several changes in biological (growth and reproductive behaviour) and distribution patterns of oilsardines along the SEAS during the period.

The present study depicts the bloom dynamics of O. magnificus in relation to water quality, phytoplankton ecology and fishery (especially oilsardine fishery) of nearshore waters off Dakshina Kannada. The unusual increase in the dinoflagellate population density was in response to the El Nino induced high-temperature and high-saline and nutrient deficient (Nitrogen limited) conditions which were favourable for the growth of mixotrpohic Ornithocercus sp. Changes in the dinoflagellate composition with the dominance of Ornithocercus sp. at high densities and its relation to

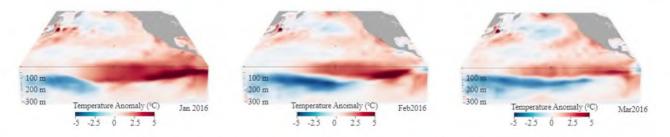


Fig. 9. Temperature anomalies in the ocean depths during the pre-bloom, bloom and post-bloom periods (by Joshua Stevens, using data from the Global Data and Assimilation Office (https://earthobservatory.nasa.gov/features/ElNino)

Lavanya Ratheesh et al.

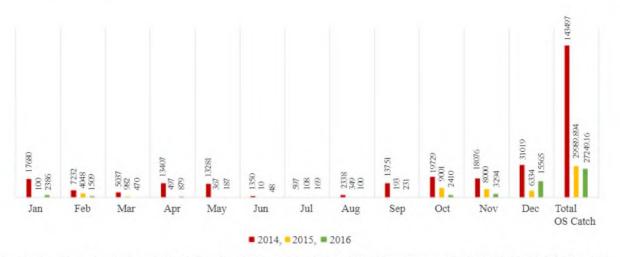


Fig. 10. Monthly and annual catch (t) of oilsardine for Karnataka during 2014-2016 (Source: NMFDC, ICAR-CMFRI, Kochi)

increasing SST have been reported along the coastal waters off Korean Peninsula (Kim et al., 2008; Kim et al., 2013) and from off Kochi coastal waters (Lavanya et al., 2020). The present bloom lasted for a month (peak abundance observed 17 days from pre-bloom phase) and caused notable changes in the water quality and phytoplankton composition. Although there was no fish mortality corresponding to the bloom event, such frequent occurrence of a rarely blooming species in the surface waters and its link to the El Nino and warmer waters indicates visible changes in the marine ecosystem. Hence, warm and high saline waters induced by strong El Nino. creates oligotrophic conditions and delayed upwelling along the SEAS, thereby favouring the dominance of dinoflagellates like O. magnificus in association with ectosymbionts over other phytoplankton species. As surface feeders like oilsardine depends on these dinoflagellates for food during the warm seasons, surface blooms of this species need to be monitored and reported.

Acknowledgments

The authors are thankful to the Director, ICAR-CMFRI, Kochi, Dr. P. Kaladharan, former Head, Fishery Environment Management Division, ICAR-CMFRI, Kochi and late L. K. Suvarna, Mangalore Regional Centre of ICAR-CMFRI for their support for the completion of this study.

Reference

- APHA 1981. Standard methods for the examination of water and wastewater. American Public Health Association, American Water Works Association and Water Pollution Control Federation, Washington, DC, USA, 1198 pp.
- Bo, L., HongLi, R., Adam, A. S., Jie, W., Nick, D., Doug, S., Jianghua, W., Rosie, E., Craig, M. and Margaret, G. 2017. An extreme negative Indian Ocean Dipole event in 2016: Dynamics and predictability. *Clm Dyn*, 51: 89-100.

- Cleve, P. T. 1900. Notes on some Atlantic plankton organisms. K. Svenska Vetensk Akad. Handl., 34(1): 1-22.
- Cleve, P. T. 1901. Plankton from the Indian Ocean and the Malay Archipelago. K. Svenska Vetensk-Akad. Handl., 35(5): 8-58.
- Cleve, P. T. 1903. Report on plankton collected by Mr. Thoruld Wulff during a voyage to and from Bombay. Ark. Zool., 1: 329-391.
- David, V. A. and William, B. W. 1960. The effect of salinity on growth of Gymnodinium breve Davis, Biolo, Bull., 119(1): 57-64.
- Dae-il, K., Yukihiko, M., Sou, N., Mineo, Y., Yang-Ho, Y., Yuji, O., Nobuyoshi, I. and Tsuneo, H. 2004. Effects of temperature, salinity and irradiance on the growth of the harmful red tide dinoflagellate *Cochlodinium polykrikoides* Margalef (Dinophyceae), J. Plankton Res., 26: 61-66. doi: 10.1093/plankt/fbh001.
- Dhulkhed, M. H. 1972. Food of the oil sardine taken by bottom nets and surface gillnet in the Mangalore area. *Indian* J. Fish., 19(1&2): 163-166.
- Droop, M. R. 1974. Heterotrophy of carbon. In: Stewart, W. D., (Ed.), Algal physiology and biochemistry. University of California Press, Barkely, USA, p. 530-599.
- Farnelid, H., Tarangkoon, W., Hansen, G., Hansen, P. J. and Riemann, L. 2010. Putative N2-fixing heterotrophic bacteria associated with dinoflagellate-*Cyanobacteria* consortia in the low-nitrogen Indian Ocean, *Aquat. Microb. Ecol.*, 61: 105-117. DOI:10.3354/ame01440.
- Gomez, F. 2005. A list of free -living dinoflagellate species in the world's oceans. Acta. Bot. Croat., 64(1): 129-212.
- Gordon, N., Angel, D. L., Neori, A., Kress, N. and Kimor, B. 1994. Heterotrophic dinoflagellates with symbiotic cyanobacteria and nitrogen limitation in the Gulf of Aqaba. *Mar. Ecol. Prog. Ser.*, 107: 83-88. DOI:10.3354/meps107083.
- John, A. G., Dionysios, E. R., George, K. and Ibrahim, H. 2018. Impacts of warming on phytoplankton abundance and phenology in a typical tropical marine ecosystem. *Sci. Rep.*, 8: 2240.

Occurrence of Ornithocercus magnificus blooms along the south-west coast of India

- Jyothibabu, R., Madhu, N. V., Maheswara, P. A., Ashadevi, C. R., Balasbramanian, T., Nair, K. K. C. and Achuthankutty, C. T. 2006. Environmentally-related seasonal variation in symbiotic associations of heterotrophic dinoflagellates with cyanobacteria in the western Bay of Bengal. *Symbiosis*, 42: 51-58.
- Kagwade, P. V. 1964. The food and feeding habits of the Indian oil sardine, Sardinella longiceps Valenciennes. Indian J. Fish., 11A(1): 345-370.
- Karthik, R., Anandavelu, I., Jayalakshmi, T., Uma, V. S., Hariharan, G., Robin, R. S., Debasitudu, Yogeshwari, S., Purvaja, R. and Ramesh, R. 2017. Ornithocercus magnificus Stein, 1883 - A bloom forming microalgae encountered from the West coast of India. In: Book of Abstracts, Fourth Indian Biodiversity Congress (IBC 2017). 10-12 March 2017, Pondicherry University Convention Centre, Puducherry.
- Kim, H. S., Jung, M. M. and Lee, J. B. 2008. The Korean Peninsula warming based on appearance trend of tropical dinoflagellate species, Genus Ornithocercus. The Sea, 13(4): 303-307.
- Kim, H. S., Kim, S. H., Jung, M. M. and Lee, J. B. 2013. New record of dinoflagellates around Jeju Island. J. Ecol. Env., 36(4): 273-291.
- Kripa, V., Mohamed, K. S., Koya, K. P. S., Jeyabaskaran, R., Prema, D., Padua, S., Kuriakose, S., Anilkumar, P. S., Nair, P. G., Ambrose, T. V., Dhanya, A. M., Abhilash, K. S., Bose, J., Divya, N. D., Shara, A. S. and Vishnu, P. G. 2018. Overfishing and climate drives changes in biology and recruitment of the Indian oil sardine *Sardinella longiceps* in South-eastern Arabian Sea. *Front. Mar. Sci.* 5: 443.
- Kofoid, C. A. and Skogsberg, T. 1928. The Dinophysidae. Mem. Mus. Comp. Zool. Harv., 51: 1-766.
- Kofoid, C. A. 1907. New species of dinoflagellates. Bull. Mus. Comp. Zool. Harv., 50: 161-208.
- Kuzmenko, L. V. 1975. Systematic composition of phytoplankton of Arabian Sea Oceanology, 34: 15-38.
- Li, W. K. 1995. Composition of ultraphytoplankton in the central North Atlantic. Mar. Ecol. Prog. Ser., 122: 1-8.
- Lucas, I. A. N. 1991. Symbionts of the tropical Dinophysiales (Dinophyceae). Ophelia, 33: 213-224. https://doi.org/10.1 080/00785326.1991.10429712.
- Lavanya, R., Reena, V. J., Parvathy, R., Shelton, P., Prema, D., Kripa, V. and Kaladharan, P. 2020. First report of a rare bloom of *Ornithocercus magnificus*, Stein 1883 along the coastal waters of Kochi; A possible indicator of increasing sea surface temperature. J. Mar. Biol. Ass. India, 62(2), 92-98.
- Lewis, E. J. 1965. On a Gonyaulax bloom off Mt. Dalley, in the Arabian Sea.[In: Krishnamurthy, V. (Ed.), Proceeding of the Seminar on sea, salt and plants. -Central Salt and Marine. Chemicals Research Institute, Bhavnagar, Gujarat, India, p. 224-226.
- Maranon, E., Behrenfeld, M. J., Gonzalez, N., Mourino, B., Zubkov, M. V. 2003. High variability of primary production in oligotrophic waters of the Atlantic Ocean: Uncoupling

from phytoplankton biomass and size structure. Mar. Ecol. Prog. Ser., 257: 1-11.

- Paerl, H. W., Kenneth, M., Crocker, M. and Prufert, L. E. 1987. Limitation of N₂ fixation in coastal marine waters: Relative importance of molybdenum, iron, phosphorus and organic matter availability. *Limnol. Oceanogr.*, 32: 525-536. https:// doi.org/10.4319/lo.1987.32.3.0525.
- Raymont, J. E. G. 1963. Plankton and productivity in the oceans. Pergamon Press, Oxford, UK, 660 pp..
- Racault, M. F., Sathyendranath, S., Menon, N. and Platt, T. 2017. Phenological responses to ENSO in the global oceans. Surv Geophys 38: 277-293. doi: 10.1007/s10712-016-9391-1.
- Rohit, P., Sivadas, M., Abdussamad, E. M., Margaret, M. R. A., Koya, K. P. S., Ganga, U., Ghosh, S., Rajesh, K. M., Mohammed Koya, K., Anulekshmi Chellappan, Mini, K. G., Grinson George, Subal Kumar Roul, Surya, S., Sandhya Sukumaran, Vivekanandan, E., Retheesh, T. B', Prakasan, D., Satish Kumar, M., Mohan, S., Vasu, R. and Supraba, V. 2018. Enigmatic Indian oil sardine: An insight.. CMFRI Special Publication No. 130. ICAR-Central Marine Fisheries Research Institute, Kochi, 156 pp.
- Saji, N. H., Goswami, B. N. and Vinayachandran, P. N. 1999. A dipole mode in the tropical Indian Ocean. *Nature*, 401: 360-363. DOI:10.1038/43854.
- Shannon, C. E. and Weaver, W. 1949. The mathematical theory of communication. University of Illinois Press, Urbana, Illinois, USA, 125 pp.
- Shetye, S. S., Siby, K., Mangesh, G. and Pottekkatt, J. V. 2019. 2015-16 ENSO contributed reduction in oil sardines along the Kerala coast, south-west India. *Mari. Ecol.*, 40: 6. https://doi.org/10.1111/maec.12568.
- Strickland, J. D. H. and Parsons, T. R. 1968. A practical handbook of seawater analysis. Bulletin of Fisheries Research Board of Canada, 167, 311 pp.
- Subrahmanyan, R. 1958. Phytoplankton organisms of the Arabian Sea off the west coast of India, *Indian Bot. Soc.*, B. 37: 435-441.
- Subrahmanyan, R. 1968. The Dinophyceae of the Indian Seas. Part I. Genus Ceratium Schrank. Mem. II. Marine Biological Association of India, Kochi, Kerala, India, 129 pp.
- Subrahmanyan, R. 1959. Studies on the phytoplankton of the west coast of India Part I. Quantitative and qualitative fluctuation of the total phytoplankton crop, the zooplankton crop and their interrelationship, with remarks on the magnitude of the standing crop and production of matter and their relationship to fish landings. *Proc. Indian Acad. Sci.*, 50(3(B)): 113-187.
- Syed, M. S., Gul, S. K. and Mahwish, K. 2008. The dinoflagellae genus Ornithocercus Stein from North Arabian Sea Shelf of Pakistan. Pak. J. Bot. 40(2): 849-857.
- Taylor, F. J. R. 1976. Dinoflagellates from International Indian Ocean Expedition. *Biblioth. Bot.*, 132: 1-234.

Lavanya Ratheesh et al.

- Tarangkoon, W., Hansen, G. and Hansen, P. J. 2010. Spatial distribution of symbiont bearing dinoflagellates in the Indian Ocean in relation to oceanographic regimes. *Aquat. Microbial Ecol.*, 58: 197-213. DOI:10.3354/ame01356.
- Tasaduq, H. S., Chakraborty, S. K., Tarkeshwar, K., Sadawarte, R. K. and Sandhya, K. M. 2019. Food and feeding habits of oil sardine Sardinella longiceps from Ratnagiri coast. Indian J. Geo-Mar., Sci., 48(3): 309-318.
- USEPA. 2012. National Coastal Condition Report IV.EPA-842-R-10-003. Office of Research and Development and Office of Water, U. S. Environmental Protection Agency, Washington, DC, USA, 334 pp.
- Weare, B. C. 1979. A statistical study of the relationships between ocean surface temperatures and the Indian monsoon. J. Atmos. Sci., 36: 2279-2291.
- Wood, E. J. F. 1954. Dinoflagellates of Australian region. Austr. J. Mar. Freshwat. Res., 5(2): 171-351.
- Wood, E. J. F. 1963. Check list of the dinoflagellates recorded form the Indian Ocean Rep. Div. Fish. Oceanogr. Commonw. Sci. ind. Res. Org. Aust., 28: 1-55.
- Wood, E. J. F. 1968. Dinoflagellates of the Caribbean Sea and adjacent areas. University of Miami Press, Coral Gables, Florida, USA, 143 pp.

Date of Receipt : 28.09.2021 Date of Acceptance : 25.08.2022